

# MAMMALS ON ICE : HIBERNATION



[www.carleton.ca/~kbstorey](http://www.carleton.ca/~kbstorey)

## Model Hibernators

*Spermophilus tridecemlineatus*,  
13-lined ground squirrel



*Spermophilus richardsonii*,  
Richardson's ground squirrel



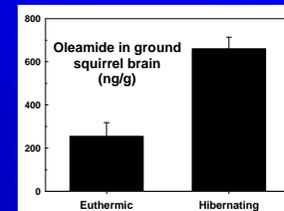
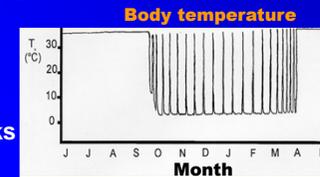
*Myotis lucifugus*, little brown bat



- Seasonal phenomenon
- Pre-hibernation hyperphagia
- Gain up to 40% of body mass
- Need polyunsaturated fats
- Find hibernaculum: dark, near 0°C

## What happens?

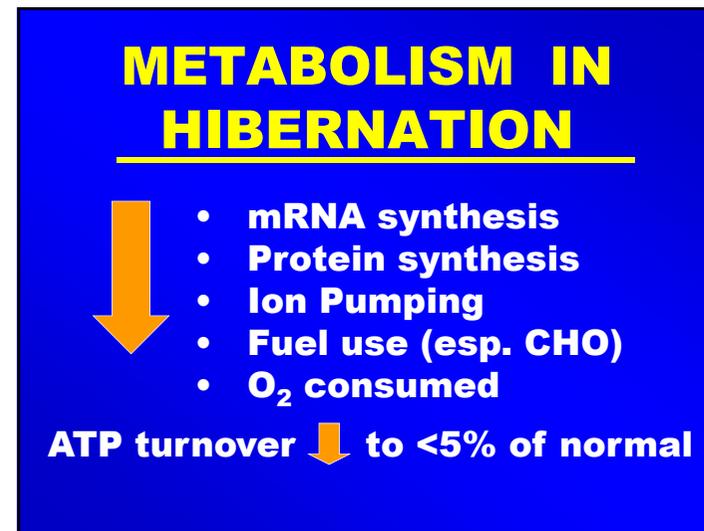
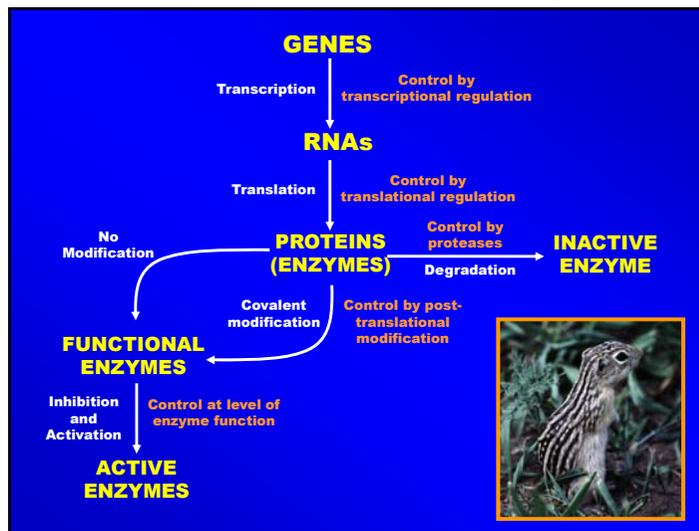
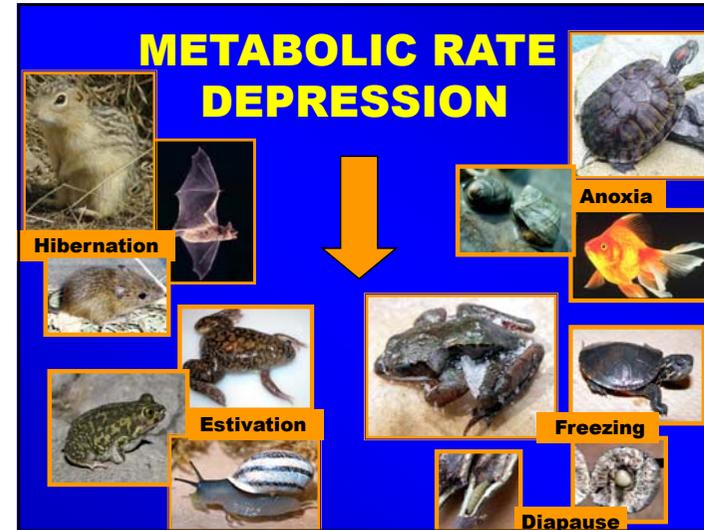
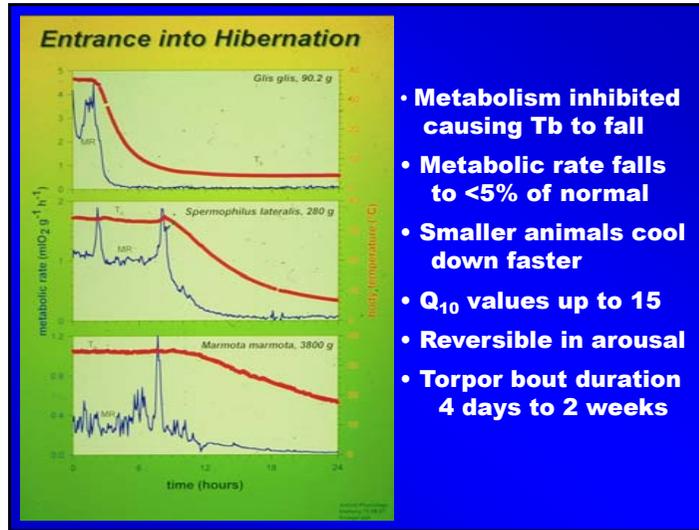
- drop in body temperature
- reduced heart rate
- apnoic breathing
- some muscle atrophy
- periods of torpor lasting weeks
- non-REM sleep
- oleamide increases in brain



- suppression of carbohydrate oxidation
- RQ of 0.7 = lipid oxidation

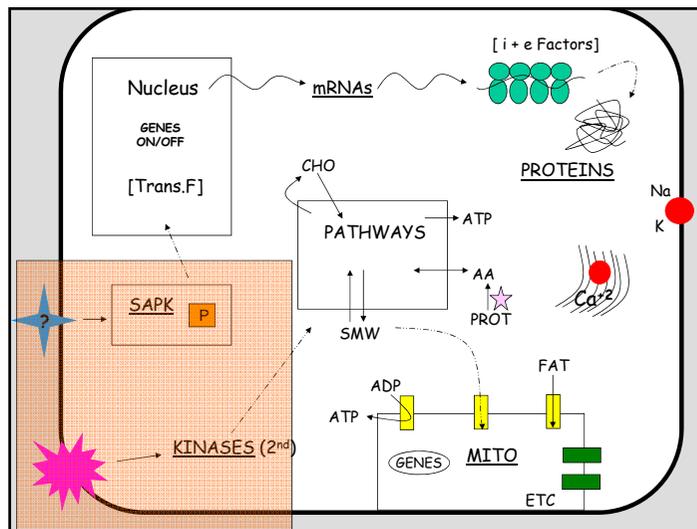
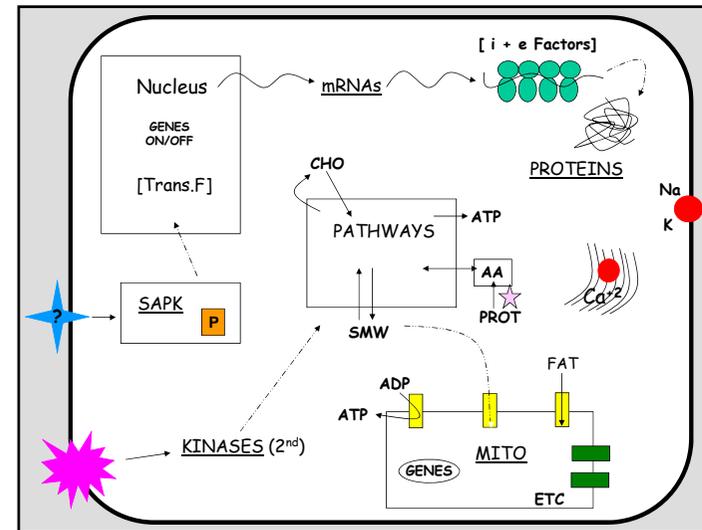
**MR falls to fraction of normal**

Stewart JM, Boudreau NM, Blakely JA & Storey KB. 2002. *J. Thermal Biol.* 27, 309-315.



# PRINCIPLES OF HIBERNATION

1. Metabolic rate reduction
2. Control by protein kinases (SAPKs, 2<sup>nd</sup> messenger PKs)
3. Selective gene activation



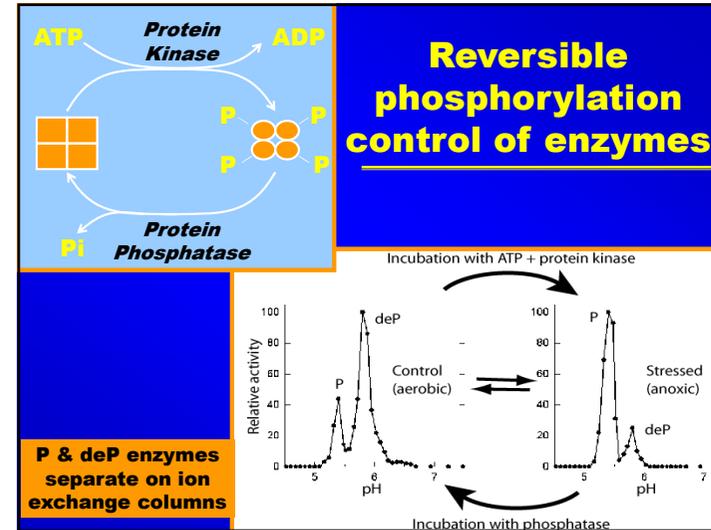
## HIBERNATION INDUCED CHANGES

- Protein Synthesis slows to 1%
- Pumps & Channels closed
- Energy Production slows to 5%
- Energy Utilization slows to 2%
- Few 'SAP' kinases activated
- Gene 'inactivation' (↓ mRNA)
- Few Genes activated (1-2%)

## PROTEIN KINASES



- Covalent modification by phosphorylation
- Families of protein kinases: PKA (cAMP), PKG (cGMP), CaM (Ca<sup>2+</sup>), PKC (Ca<sup>2+</sup>, PL, DG)
- SAPKs : daisy chain phosphorylations
- Regulation via interconversion of active vs subactive forms of protein substrates
- p38, ERK (1/2), JNK, AMPK, AKT (mTOR)



## PATHWAY CONTROL IN HIBERNATION

### Phospho / de-Phospho

- Glycolysis (GP, GS, PFK, PK)
- Fat synthesis (ATP-CL, ACC)
- CHO fuel use (PDH)
- Translation (eIF2 $\alpha$ , eEF2)
- Ion pumps (NaK, Ca-ATPase)
- *the usual suspects, TextBook*



1. Novel PhosphoEnzymes: BioInformatics + Phospho-analyses
2. <sup>32</sup>P-ATP labeling studies
3. Purification / Properties
4. Structure / Function
5. Phospho-sites

## Post-translational Modifications: The Next Generation

### Novel Phosphorylation Control

CK, GDH, Hexokinase, G6PDH,  
LDH, NADP-IDH,  $\alpha$ -GPDH, AMPD,  
GAPDH, FBPase, Antioxidant enzymes

**PTM:** Acetylation, Methylation,  
SUMOylation

## HIBERNATION INDUCED CHANGES

- **Protein Synthesis slows to 1%**
  - **Pumps & channels closed**
  - **Energy Production slows to 5%**
  - **Energy Utilization slows to 2%**
  - **Few 'SAP' kinases activated**
- **Gene 'inactivation' ( ↓ mRNA )**
  - **Few Genes activated (1-2%)**

## TURNING OFF GENES: Role of Epigenetics

### Epigenetics:

- **Stable changes in gene activity that do not involve changes in DNA sequence**

### Common mechanisms:

- **DNA methylation**
- **Histone modification / histone variants**  
e.g. **acetylation, phosphorylation**
- **Regulatory non-coding RNAs**

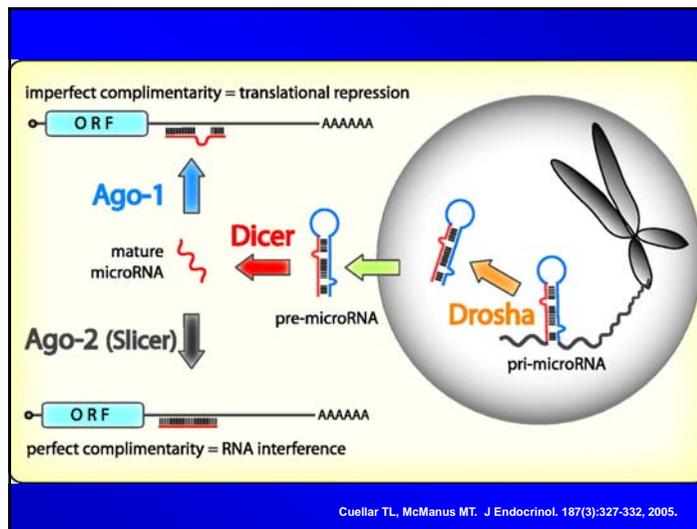


## Transcription Suppression in Hibernator Muscle

- Phospho-Histone H3 (Ser10) levels reduced
- Acetyl-Histone H3 (Lys23) levels reduced  
\* **Both inhibit Transcription** \*
- Histone Deacetylase activity increased 80%
- HDAC 1 & 4 protein levels increased
- RNA Polymerase II activity Decreased

## Regulatory non-coding RNAs microRNA

- Small RNAs ~22 nucleotides in length
- Highly conserved across species
- Bind to 3' UTR of mRNAs
- Could be 1000, affect 60 % of genes
- Disease involvement
- Act to :
  - Block translation of mRNA
  - Target mRNA for degradation



## Are miRNAs differentially regulated in hibernators?

- **Yes!** Selected miRNAs were regulated in heart, muscle & kidney of hibernating 13-lined ground squirrels

(Morin, Dubuc & Storey, 2008, Biochim Biophys Acta 1779:628-633)

miRNA	Fold change	Process in higher vertebrates
Mir-1	2.0	Myogenesis
Mir-133a	2.4	Myogenesis
Mir-206	2.6	Myogenesis
Let-7	2.0	Cell cycle
Mir-26	2.4	Hypoxia
Mir-451	2.6	Erythropoiesis

## Turning it all off

Journal of Molecular Cell Biology Advance Access published December 21, 2010  
doi:10.1093/jmcb/mbq004

### Review

#### The emerging roles of microRNAs in the molecular responses of metabolic rate depression

Kyle K. Biggar and Kenneth B. Storey\*

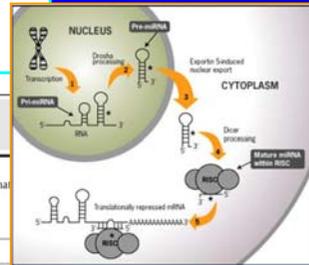
Institute of Biotechnology and Department of Biology, Carleton University, 1125 Colonel By Drive, Ottawa, ON, Canada K1S 5B4

\* Correspondence to: Kenneth B. Storey, Tel: +1 613 520 5276; Fax: +1 613 520 5245; E-mail: kenneth\_storey@carleton.ca

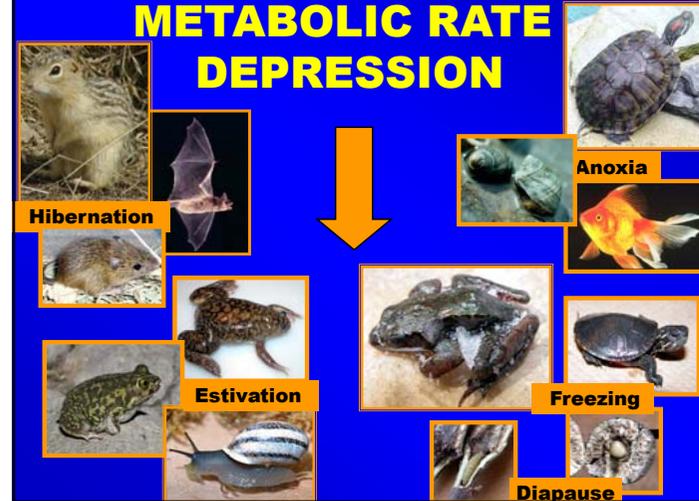
MicroRNAs, a class of small non-coding RNA molecules, have been shown to regulate gene expression in a variety of organisms and tissues. In this review, we discuss the emerging roles of microRNAs in the molecular responses of metabolic rate depression.

Keywords: hibernation, metabolic rate depression, microRNAs, transcription, translation

Journal Pre-proof



## METABOLIC RATE DEPRESSION

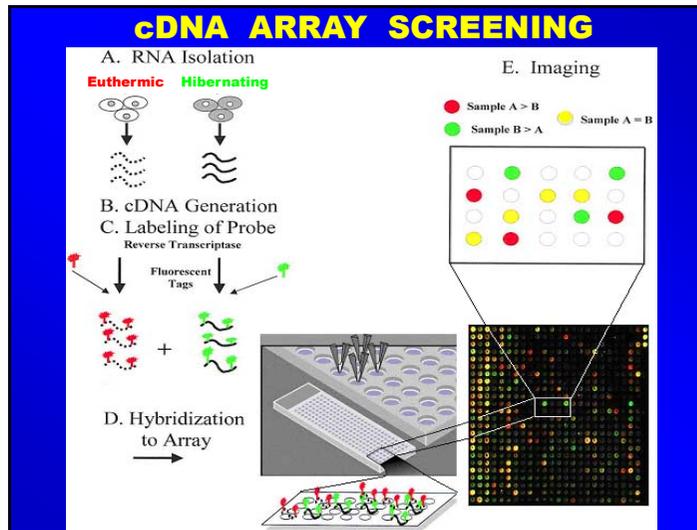


## HIBERNATION INDUCED CHANGES

- Protein Synthesis slows to 1%
- Pumps & channels closed
- Energy Production slows to 5%
- Energy Utilization slows to 2%
- Few 'SAP' kinases activated
- Gene 'inactivation' ( ↓ mRNA )
- Few Genes activated (1-2%)

## ROLE OF TRANSCRIPTION

- Global rate of mRNA synthesis depressed. Method: nuclear run-on
- Are selected genes up-regulated ?
- TO ASSESS GENE UPREGULATION:  
What new mRNAs are created  
- cDNA library, Gene Chip  
Sequenced genome(s) as of 2011



## cDNA Arrays

- Methods
- Materials
- Sources
- Publications

**Carleton University**  
 LDRS | Institute of Biochemistry | Department of Biology | Department of Chemistry |  
 Faculty of Graduate Studies and Research | Carleton University website |  
 Carleton library electronic journals | (416) 497-1111 |  
 Safety - [Matter Control](#) | [Shore Lab Chemical List](#)

**Kenneth B. Storey**  
 Ph.D., F.R.S.C.  
**Canada Research Chair in Molecular Physiology**  
 Professor of Biochemistry,  
 Institute of Biochemistry,  
 and Departments of  
 Biology and Chemistry II

Contact information:  
 Institute of Biochemistry  
 College of Natural Sciences  
 1375 Colonel By Drive  
 Ottawa, Ontario, Canada  
 K1S 5B6

Tel: +1 413 528-3628  
 Fax: +1 413 528-2569  
 Office: 508 Stacks  
 Chemistry Bldg

**Highly Cited Researcher**  
[Click to view record](#)

Email: [kenneth.storey@carleton.ca](mailto:kenneth.storey@carleton.ca)

**Diabetes** *Diabetes*, 107 Times May 2005  
*Suspended animation*, US News & World Report, May 2005  
**DISCOVER** magazine, Feb. 2005  
 Carleton U. *Insights*, Winter 2005  
 Washington Post *Science*, Dec. 2004

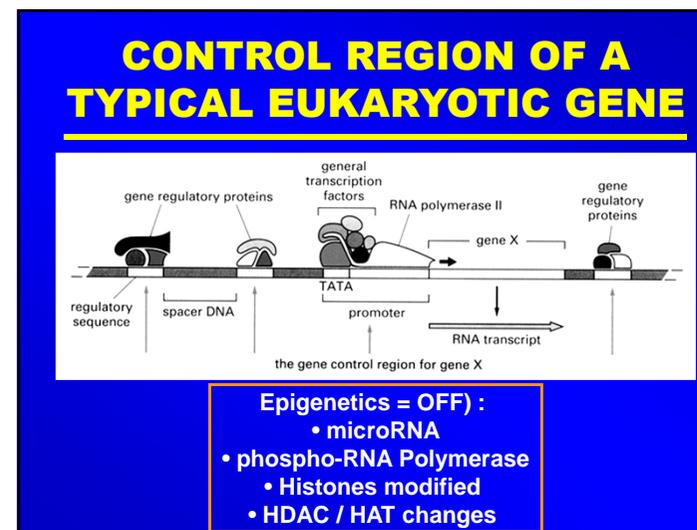
**Links:** book, magazine and newspaper articles \*\* Ken in Fiction \*\* World Free Fiction \*\* Ontario high school texts  
[Canada Research Chair Tier I](#)  
[PHOTOS - Ken's profile](#)

**THE LAB**

<a href="#">Research interests</a>	<a href="#">Professional Information</a>
<a href="#">Facilities available</a>	<a href="#">NEW Reviews &amp; Popular Articles</a>
<a href="#">Lab personnel, Past and Present</a>	<a href="#">RECENT PUBLICATIONS - 1998 - Present</a>
<a href="#">Extra scientific programs</a>	<a href="#">Publications - 1998 - 1995</a>
<a href="#">Awards - Info on running cDNA arrays</a>	<a href="#">Publications - 1974 - 1995</a>
<a href="#">PHOTO GALLERY - on animals &amp; their studies</a>	<a href="#">Book, magazine, newspaper articles &amp; Fiction</a>

## GENE CHANGES IN HIBERNATION

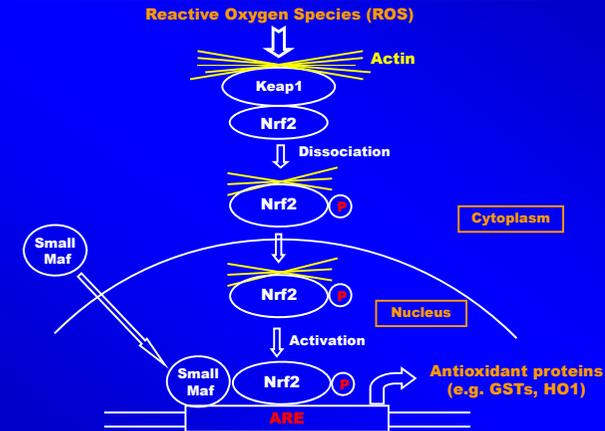
- **cDNA Library screen**
  - Mitochondrial Genes
  - AOE
  - FABP, CPT, etc.
  - Shock proteins (GRP, HSP)
  - Transcription factors
- **DNA Chip ~1-2% ↑**



## TRANSCRIPTION FACTORS

- ATF (Glucose Regulated Proteins)
- HIF (O<sub>2</sub>), HSF (Hsp)
- NFκB (IκB-P), Nrf-2 (\*\*), NRF-1
- PPAR, PGC, RXR, chREBP, CREB-P
- STAT, SMAD, p53-P, HNF, AP (1,2)
- Methods: EMSA, ChIP

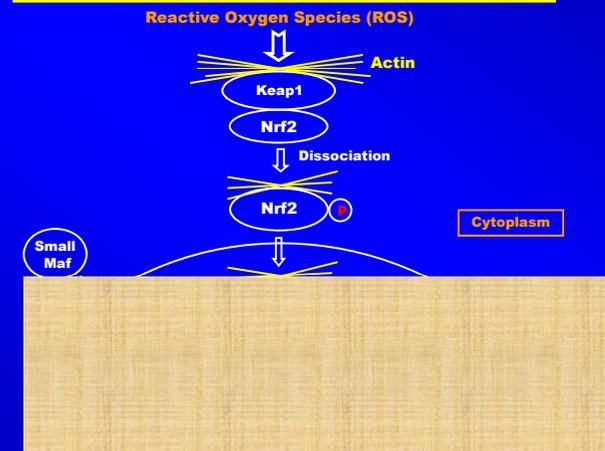
## Nrf2/ARE pathway



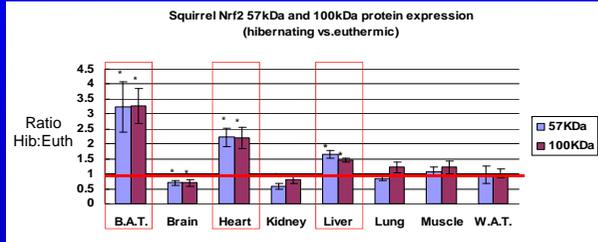
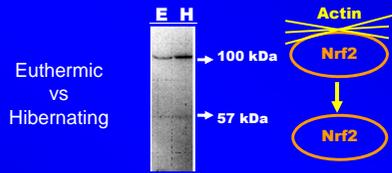
## Nrf-2

- Increased Nrf-2 protein & P
- Increased Nrf-2 in the Nucleus
- Increased levels of co-Tf: MafG
- Downstream gene activation:
- GST, HO-1, HO-2, Peroxiredoxin
- Thioredoxin, SOD (Cu/Zn & Mn)

## Nrf2/ARE pathway

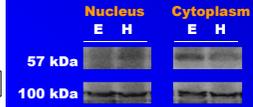
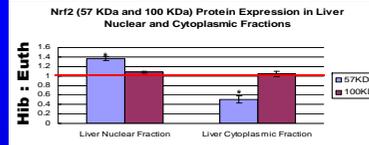


## Protein Regulation of Nrf2

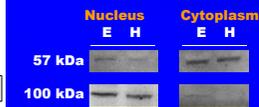
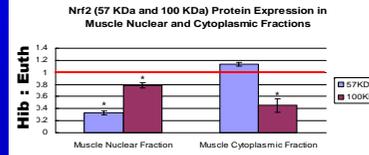


## Nrf2 distribution between nuclear and cytoplasmic fractions

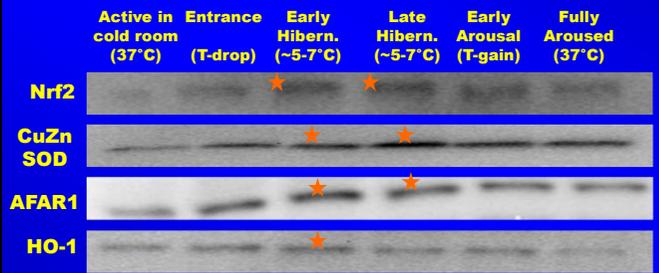
### Liver



### Muscle



## Nrf2 Timecourse in Heart

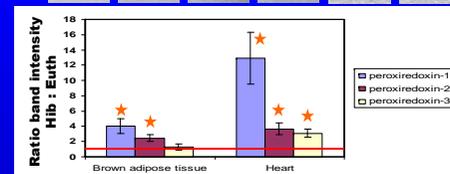


- Nrf2 protein ↑ in early and late hibernation → Up-regulation "cascade".

## Peroxiredoxins

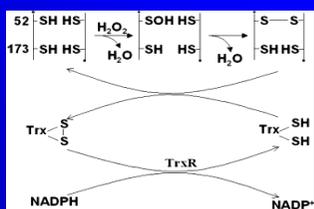
- Detoxify / reduce hydroperoxides
- Expressed at high levels
- ARE in promoter region of Prdx genes
- Nrf2 activated

### Peroxiredoxin protein levels

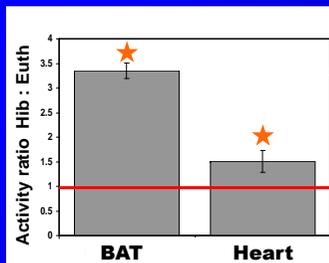


## Peroxiredoxin Activity

- Protein level correlates with increased activity
- Assays in BAT and heart with thioredoxin, thioredoxin reductase and NADPH:



Kim et al., 2005



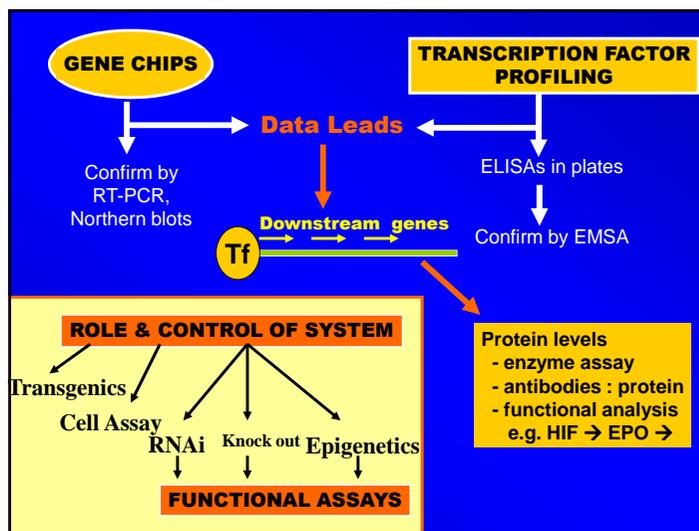
## Nrf Conclusion

### Activation of the Nrf2 pathway:

- Activated in early-late torpor, along with downstream gene protein products
- Increased PRDX, HO & TRX protein and activity

### Result:

- Detoxification of ROS, intracellular signaling control



## Where do we go from here?

- Applications of MRD research
- Novel phosphorylations
- Atrophy, hypertrophy -- autophagy for survival
- Turning it all off -- microRNA
- Epigenetics & adaptation
- Life span extension
- Antioxidant Defense
- Cell cycle suppression
- Unity through evolution



## NEW DIRECTIONS

# PRIMATE HIBERNATION!! GREY MOUSE LEMUR



# Hibernation and medicine

*Mol. Cell. Biochem.* 2010;327:77-108.  
**Metabolic rate depression: the biochemistry of mammalian hibernation.**  
 Storey KB, Storey JM  
 Institute of Biochemistry, Carleton University, Ottawa, Ontario, Canada. kenneth\_storey@carleton.ca



**Primates !!**

**Abstract**  
 During winter hibernation, small mammals fall into long periods of deep cold torpor where metabolic rate is suppressed by >90% and core body temperature can fall to near 0 degrees C. Studies with hibernators illustrate the molecular regulatory mechanisms that coordinate the suppression of metabolic functions during torpor, replete energy use, and preserve/stabilize macromolecules to support long-term viability during cold torpor. This review explores mechanisms including post-translational modification of proteins, differential regulation of enzymes, global suppression of transcription and translation including a role for microRNAs, and differential regulation of transcription factors. The review is relevant to issues in clinical medicine, such as hypothermia, and atrophy resistance.

*Decompression*, 2010;16(2):220-30. Epub 2009 Jul 14.  
**Out cold: biochemical regulation of mammalian hibernation - a mini-review.**

Storey KB  
 Institute of Biochemistry, Carleton University, Ottawa, Ont., Canada. kenneth\_storey@carleton.ca

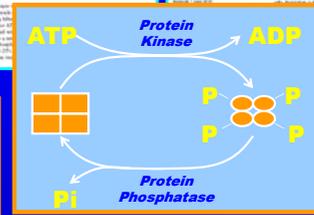
**Abstract**  
 Hibernating mammals offer an intriguing example of natural torpor and illustrate the regulatory mechanisms that control cell preservation strategies that support long-term viability in a hypometabolic state. These mechanisms include the hypothermic preservation of human organs for transplant, and guidelines that serve as an intervention strategy in human medicine. Recent advances in hibernation research contribute to metabolic depression by orchestrating the global suppression of ATP production including multiple forms of post-translational modification of proteins/enzymes (oxidation), mRNA storage mechanisms, and differential expression of microRNA species. These advances also contributed new advances in understanding the range of cell functions that are maintained during hibernation, and the implementation of the unfolded protein response, and the enhancement of mitochondrial function to control the actions of intracellular proteases in cell death and inflammation responses.



# Novel phosphorylations

Regulation of skeletal muscle creatine kinase from a hibernating mammal  
 Khalil Abuasa, Kenneth B. Storey\*  
 Institute of Biochemistry and Department of Biology, Carleton University, 112 Colonel By Drive, Ottawa, Ont., Canada K1S 5B6  
 Received 11 May 2007; accepted 15 June 2007  
 Available online 12 August 2007

Regulation of liver glutamate dehydrogenase by reversible phosphorylation in a hibernating mammal  
 Ryan AV, Bell, Kenneth B. Storey\*  
 Institute of Biochemistry and Department of Biology, Carleton University, 112 Colonel By Drive, Ottawa, Ontario, Canada K1S 5B6



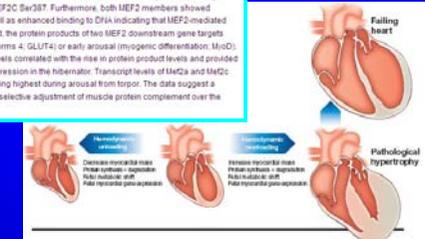
# Atrophy – Hypertrophy

*Mol. Cell. Biochem.* 2010;344(1-2):151-62. Epub 2010 Jun 9.  
**Expression of myocyte enhancer factor-2 and downstream genes in ground squirrel skeletal muscle during hibernation.**



Tasayer, GH, Storey KB  
 Institute of Biochemistry & Department of Biology, Carleton University, Ottawa, ON, Canada.

**Abstract**  
 Myocyte enhancer factor-2 (MEF2) transcription factors regulate the expression of a variety of genes encoding contractile proteins and other proteins associated with muscle performance. We proposed that changes in MEF2 levels and expression of selected downstream targets would aid the skeletal muscle of thirteen-lined ground squirrels (*Spermophilus tridecemlineatus*) in meeting metabolic challenges associated with winter hibernation, e.g., cycles of torpor-arousal, body temperature that can fall to near 0°C, long periods of inactivity that could lead to atrophy. MEF2A protein levels were significantly elevated when animals were in torpor (meanately 2.8-fold higher than in active squirrels) and the amount of phosphorylated active MEF2A Thr152 increased during entrance into torpor. MEF2C levels also rose significantly during entrance and torpor as did the amount of phosphorylated MEF2C Ser387. Furthermore, both MEF2 members showed elevated amounts in the nuclear fraction during torpor as well as enhanced binding to DNA indicating that MEF2-mediated gene expression was up-regulated in torpid animals. Indeed, the protein products of two MEF2 downstream gene targets, myocyte enhancer factor-2B (MEF2B) and myocyte enhancer factor-2C (MEF2C), were up-regulated in skeletal muscle during hibernation. MEF2B and MEF2C mRNA levels correlated with the rise in protein product levels and provided MEF2-mediated gene expression in the hibernator. Transcript levels of Mef2a and Mef2c in skeletal muscle were highest during arousal from torpor. The data suggest a gene transcription in the selective adjustment of muscle protein complement over the





# Unity through Evolution



Int. J. Biol. Sci. 2010, 6

*International Journal of Biological Sciences*  
2010, 6(1):9-20  
© Copyright International Publisher. All rights reserved.

Review

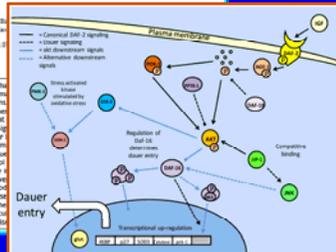
**An Overview of Stress Response and Hypometabolic Strategies in *Caenorhabditis elegans*: Conserved and Contrasting Signals with the Mammalian System**

Benjamin Lant and Kenneth B. Storey<sup>1\*</sup>  
Institute of Biochemistry, Carleton University, Ottawa, Ont., Canada

\* Correspondence to: Kenneth B. Storey, Institute of Biochemistry, Carleton University, Ottawa, Ont. K1S 5B6, Canada. Tel.: +1 613 552 3670, Fax: +1 613 552 3670  
Received: 2009/09/11, Accepted: 2009/11/25, Published: 2010/01/01

**Abstract**  
Studies of the molecular mechanisms the physiological have long been used to model organism, *Caenorhabditis elegans*, dauer stage. This period of development is metabolic rate, triggered by ambient stress. *C. elegans* employs a number of self-unfavourable conditions and survive for the suppression of cellular metabolism. The survival of nematodes through the dauer stage is fundamental to control general, mammalian systems are highly temperatures and low oxygen), however equal transcription pathways of nematode protein targets in the stress response maintained, and other differ only in the outlines a framework of critical molecules as therapeutic targets for addressing dis-

WWCeD



# HIBERNATION

- J. STOREY
  - S. EDDY
  - D. HITTEL
  - J. MacDONALD
  - A. FAHLMAN
  - P. MORIN
  - C. HOLDEN
  - H. MEHRANI
  - J. NI
- M. HAPSATOU
  - S. TESSIER
  - M. WU
  - S. BROOKS
  - C. FRANK
  - J. HALLENBECK
  - D. THOMAS
  - A. RUBTSOV
  - J. STEWART

Funded by NSERC Canada

[www.carleton.ca/~kbstorey](http://www.carleton.ca/~kbstorey)