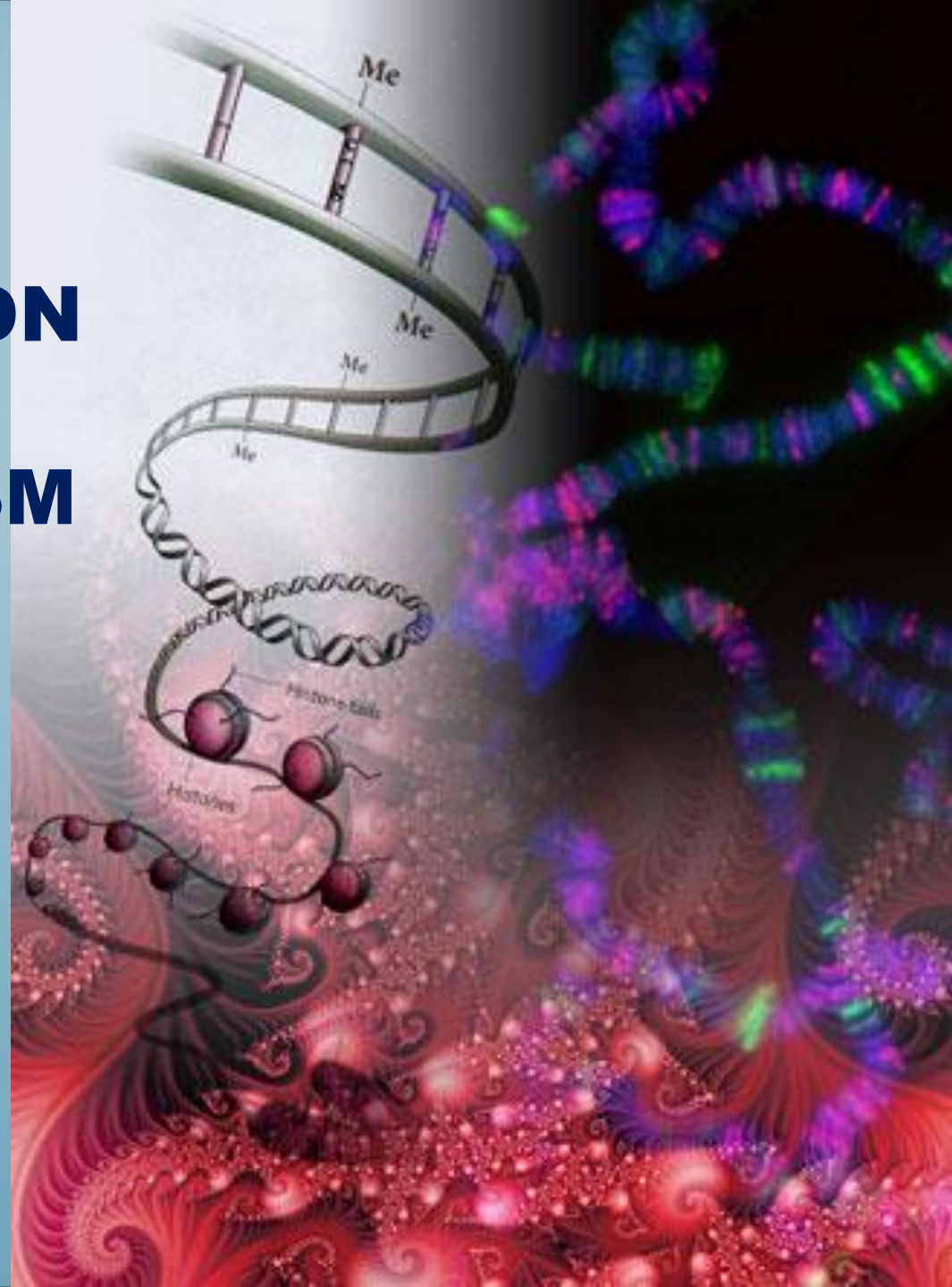


EPIGENETICS, GENE REGULATION & HYPOMETABOLISM

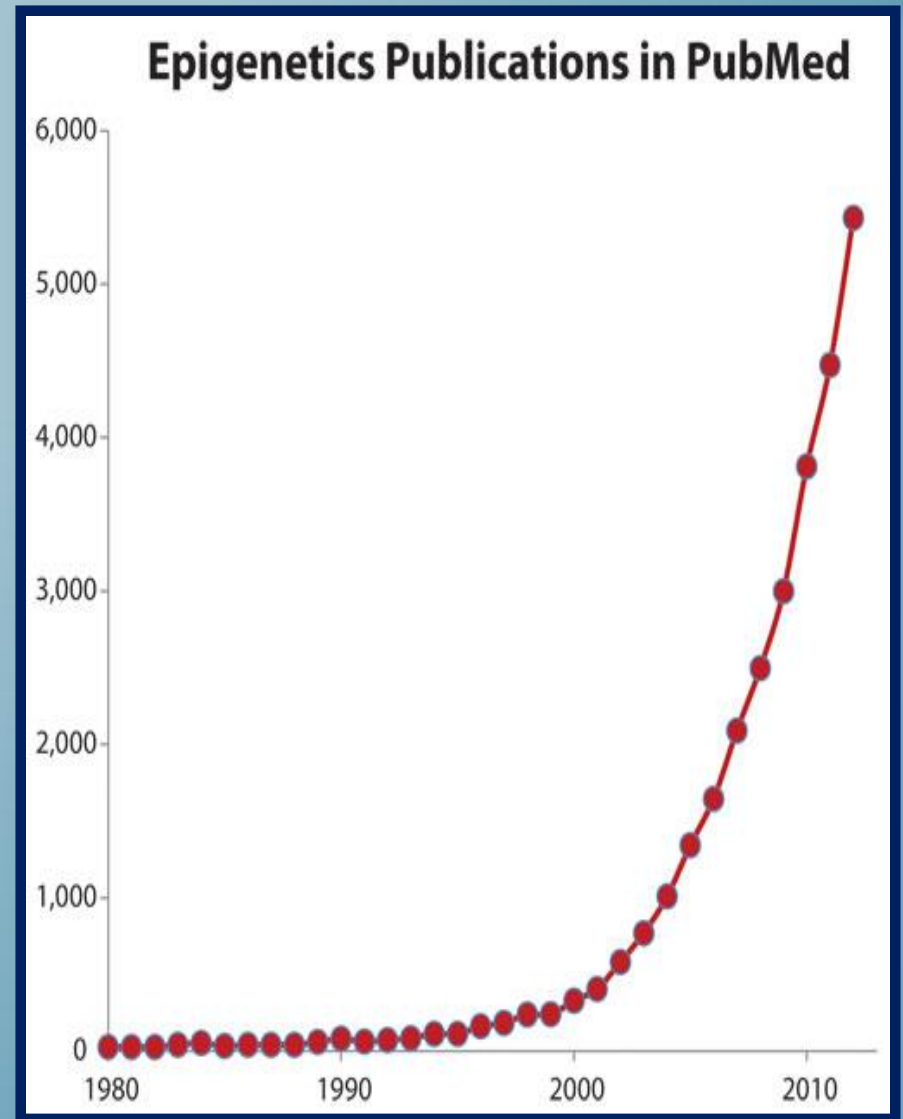
Kenneth B. Storey
Carleton University

www.carleton.ca/~kbstorey

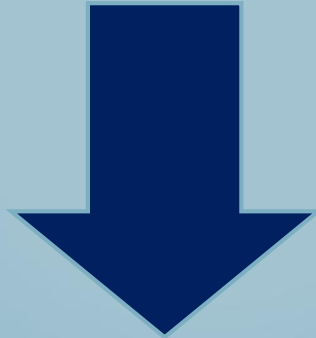


EPIGENETIC RESEARCH

The number of publications in the field increased dramatically in the last 10 years.



METABOLIC RATE DEPRESSION



Hibernation



Estivation



Anoxia



Freezing



Diapause

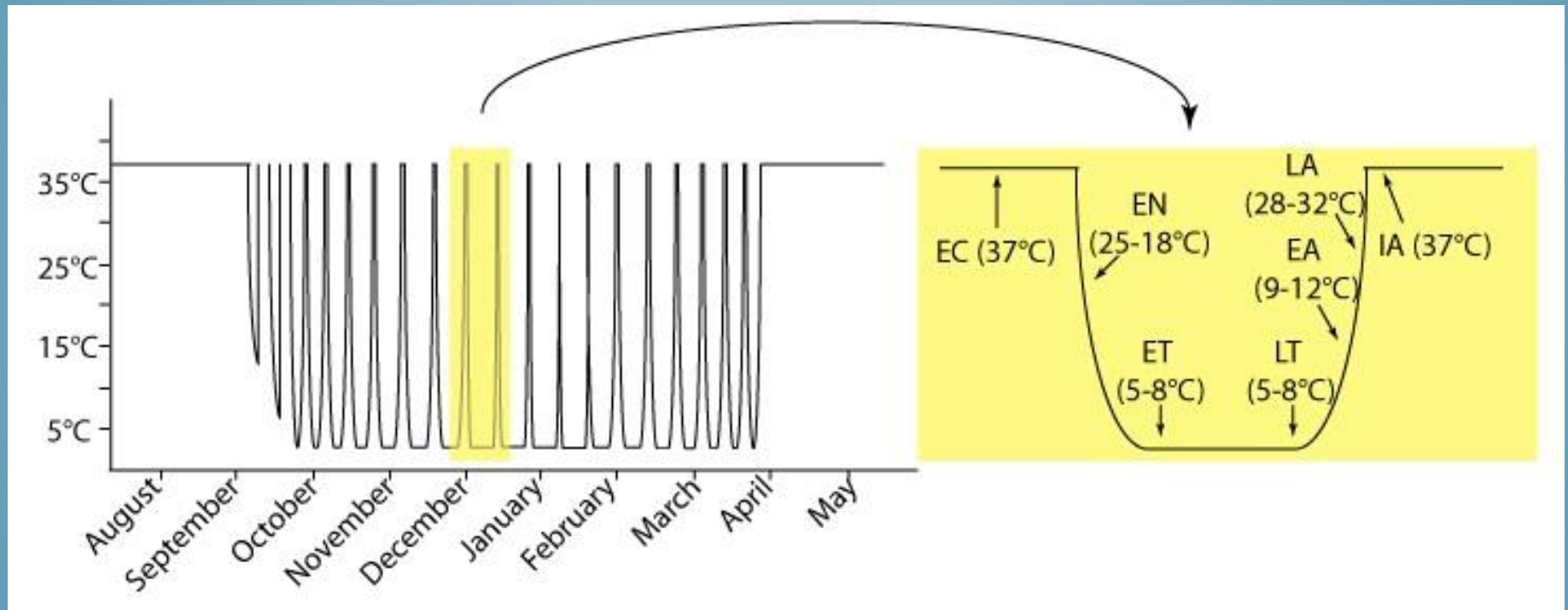


MAMMALIAN HIBERNATION

- Key characteristics :
 - metabolic rate depression (hypometabolism)
 - low body temperatures
 - Hibernation is a NATURAL model system
- Purpose is to overcome food shortages and the high energetic costs of endothermy (warm-blooded)



TORPOR-AROUSAL



Animal studies by Dr. JM Hallenbeck and Dr. DC McMullen, NIH

HIBERNATION



13-LINED GROUND SQUIRREL
Ictidomys tridecemlineatus

HIBERNATION



© Bill Kraus

Little Brown Bat, *Myotis lucifugus*



DAILY TORPOR



Grey mouse lemur, *Microcebus murinus*

ESTIVATION



Milk snail
Otala lactea

ESTIVATION



Spadefoot toad
Scaphiopus holbrookii



ANOXIA TOLERANCE



Painted turtle
Chrysemys picta



Red-eared turtle
Trachemys scripta elegans



Periwinkle
Littorina littorea

FREEZE TOLERANCE



Wood frog
Rana sylvatica

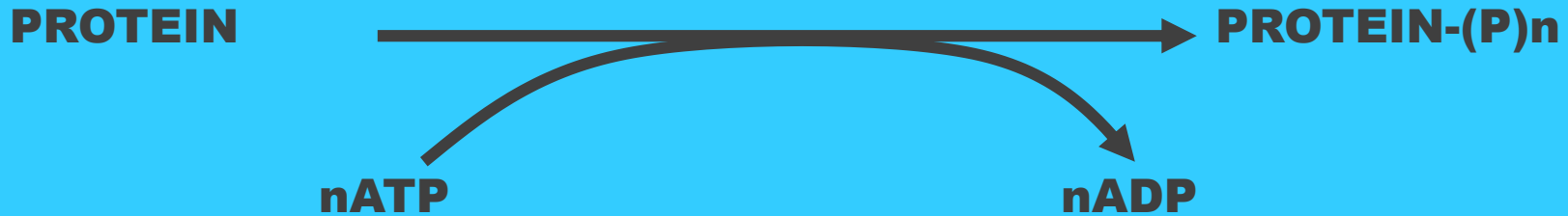
PRINCIPLES OF MRD

- 1. Metabolic rate reduction***
- 2. Control by protein kinases***
(SAPKs, 2nd messenger PKs)
- 3. Most Genes OFF***
- 4. Selective gene activation***

Same for ALL systems

www.carleton.ca/~kbstorey

PROTEIN KINASES



- Covalent modification by phosphorylation
- Families of protein kinases: PKA (cAMP), PKG (cGMP), CaM (Ca²⁺), PKC (Ca²⁺, PL, DG)
- SAPKs : daisy chain phosphorylations
- Regulation via interconversion of active vs subactive forms of protein substrates
- **p38, ERK (1/2), JNK, AMPK, AKT (mTOR)**

Phospho / de-Phospho

PATHWAY CONTROLS :

- ALL PATHWAYS, REGULATION IN MINUTES,
- REVERSED BY PROTEIN PHOSPHATASES
- METABOLIC COST = <1 % TOTAL ENERGY
- MANY NEW ENZYME TARGETS DISCOVERED

- 
- **Glycolysis** (GP, GS, PFK, PK)
 - **Fat synthesis** (ATP-CL, ACC)
 - **CHO fuel use** (PDH)
 - **Translation** (eIF2 α , eEF2)
 - **Ion pumps** (NaK, Ca-ATPase)

PRINCIPLES OF MRD

1. Metabolic rate reduction

2. Control by protein kinases
(SAPKs, 2nd messenger PKs)

3. Most Genes OFF

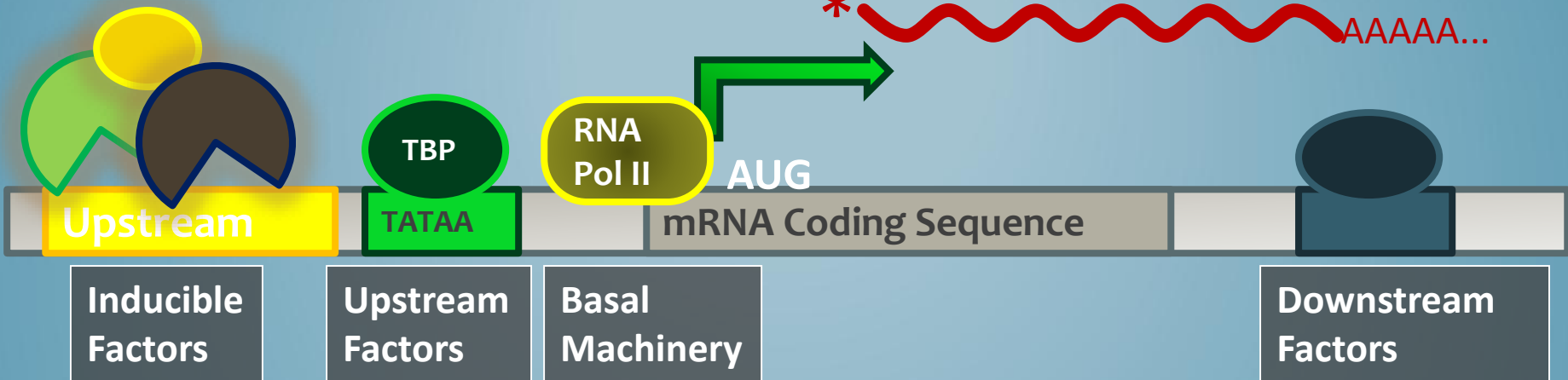
4. Selective gene activation

Same for ALL systems

Regulation of Gene Transcription

Transcription Factors

Transcription



10

Beyond gene chips: transcription factor profiling in freeze tolerance

KENNETH B. STOREY

Institute of Biochemistry, Carleton University, Ottawa, Canada K1S 5B6; kenneth_storey@carleton.ca

Abstract

The Wood Frog, *Rana sylvatica*, is one of several terrestrially hibernating anurans that display natural freeze tolerance. The multifaceted biochemical responses to the cellular stresses imposed when ~65% of total body water is converted to extracellular ice have

TRANSCRIPTION FACTORS

- *ATF (Glucose Regulated Proteins)*
- *HIF (O₂), HSF (Hsp)*
- *NFkB (Ikb-P), Nrf-2, NRF-1*
- *PPAR, PGC, RXR, chREBP, CREB-P*
- *STAT, SMAD, p53-P, HNF, AP (1,2)*

- *Methods: EMSA, CHiP*

PRINCIPLES OF MRD

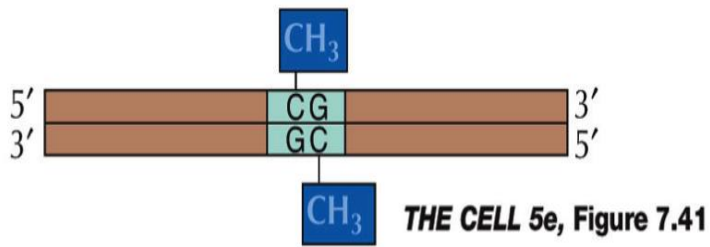
1. Metabolic rate reduction

*2. Control by protein kinases
(SAPKs, 2nd messenger PKs)*

3. Most Genes OFF -- How??

4. Selective gene activation

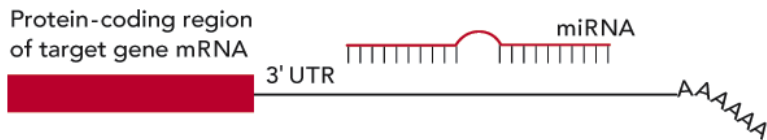
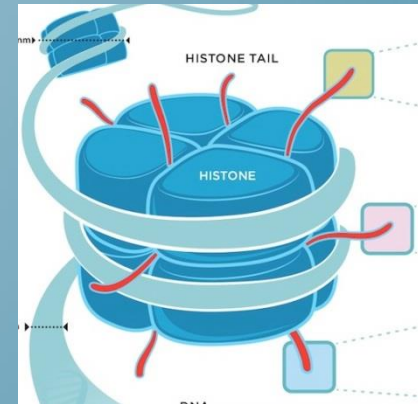
Same for ALL systems



1. DNA Methylation. Methylation of cytosines at CpG dinucleotides in promoter regions. Methylation attenuates gene expression.

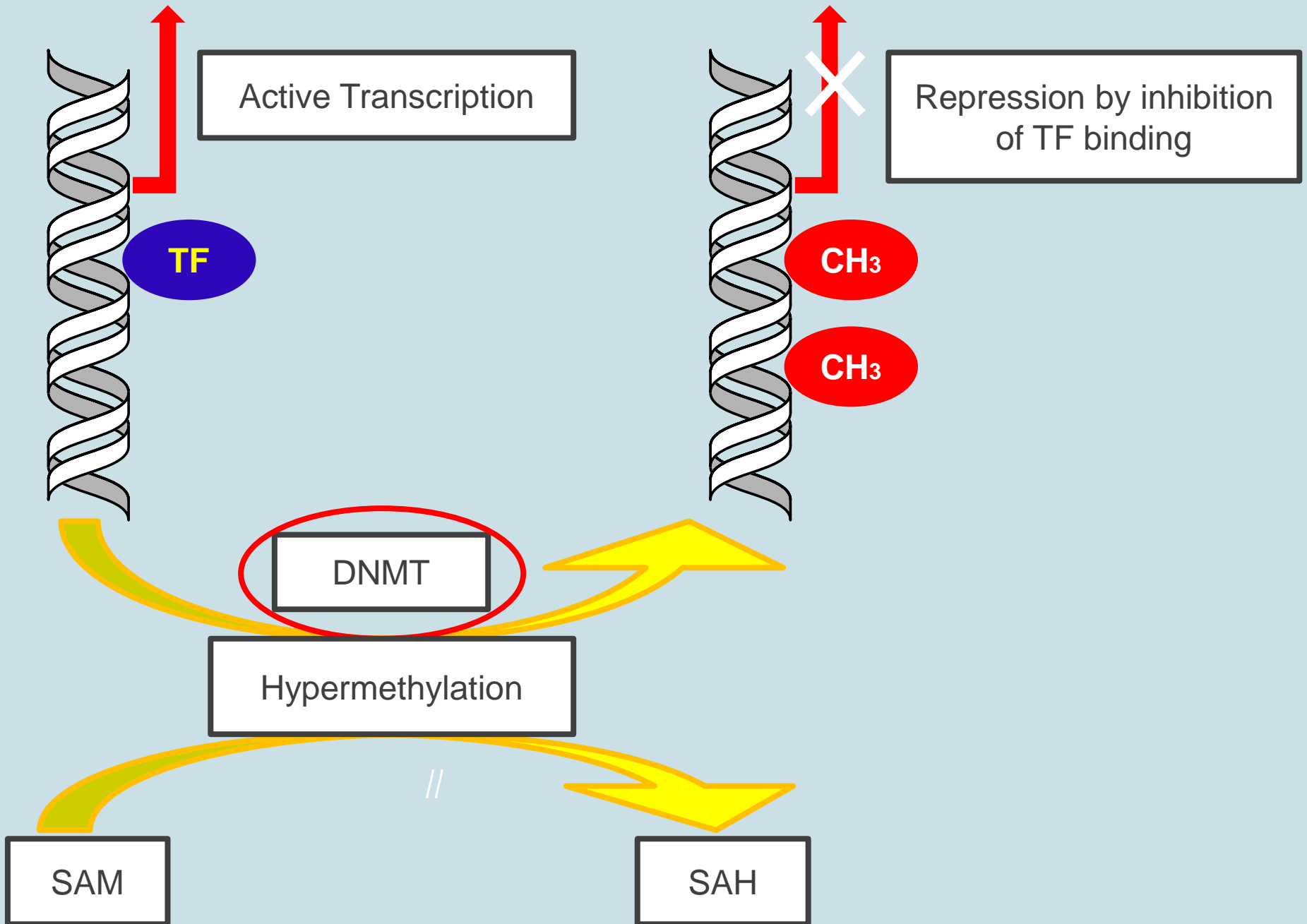
Epigenetics refers to the study of heritable changes in gene expression that are not dependent on gene DNA sequence.

2. Histone Modification. Post-translational modifications on histone tails affect histone:DNA interactions to influence accessibility of promoter regions to transcriptional machinery.

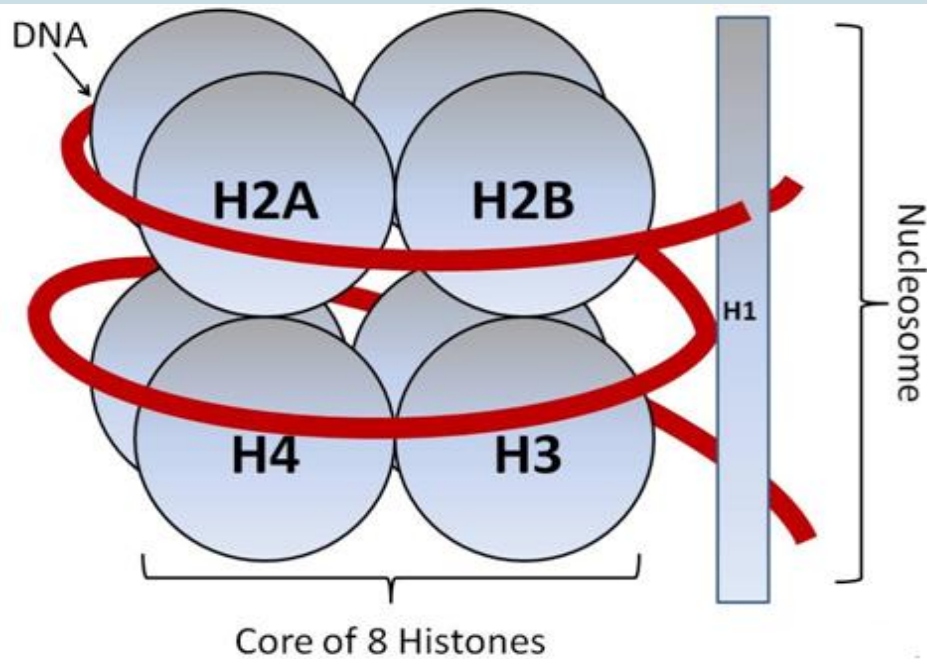


3. Non coding RNAs. microRNAs base-pair with complementary sequences in mRNA to achieve translational repression or target degradation

DNA METHYLATION



HISTONE MODIFICATION

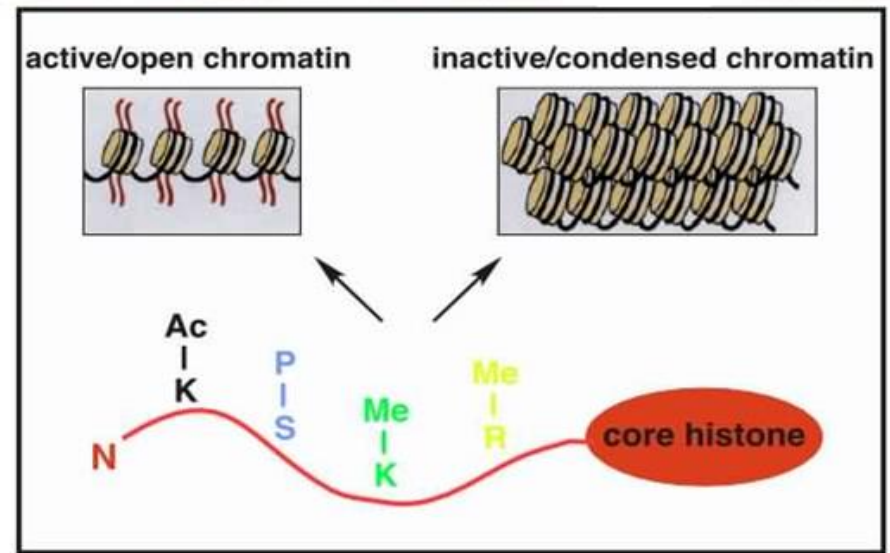
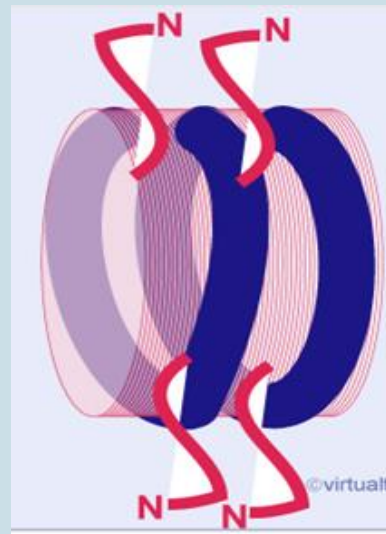


Nucleosome:

- Made of 8 histone proteins (Octamer) + linker protein
- Chromatin wraps around the histones (“beads on a string”)

3 main types of modifications:

- Methylation
- Acetylation
- Phosphorylation



Higher compaction = Lower expression

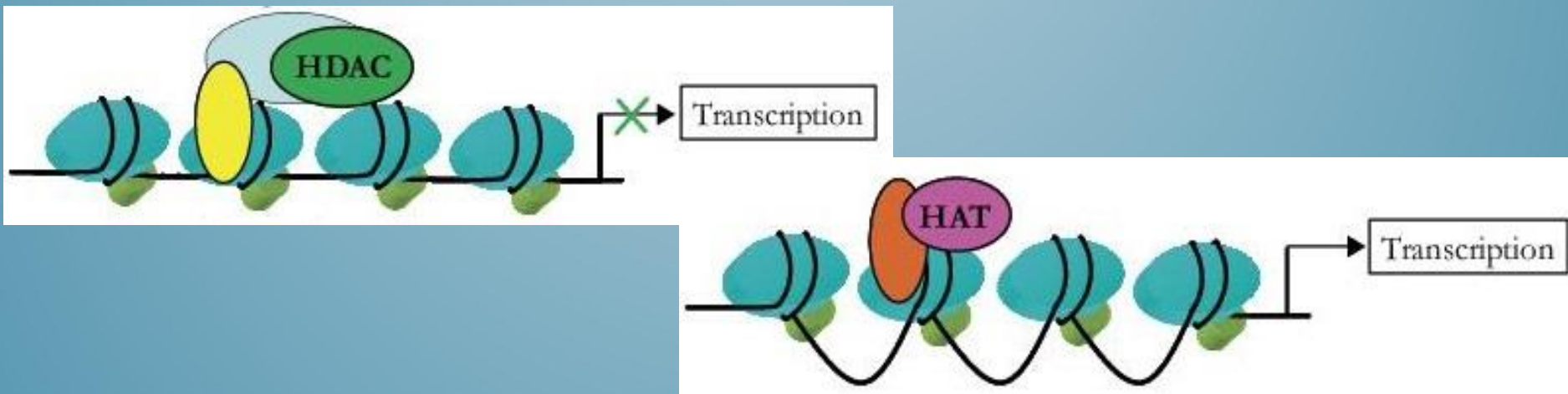
THE “HISTONE CODE”

This code is maintained by:

“WRITERS,” enzymes that can methylate and acetylate

“ERASERS,” enzymes that can demethylate and deacetylate

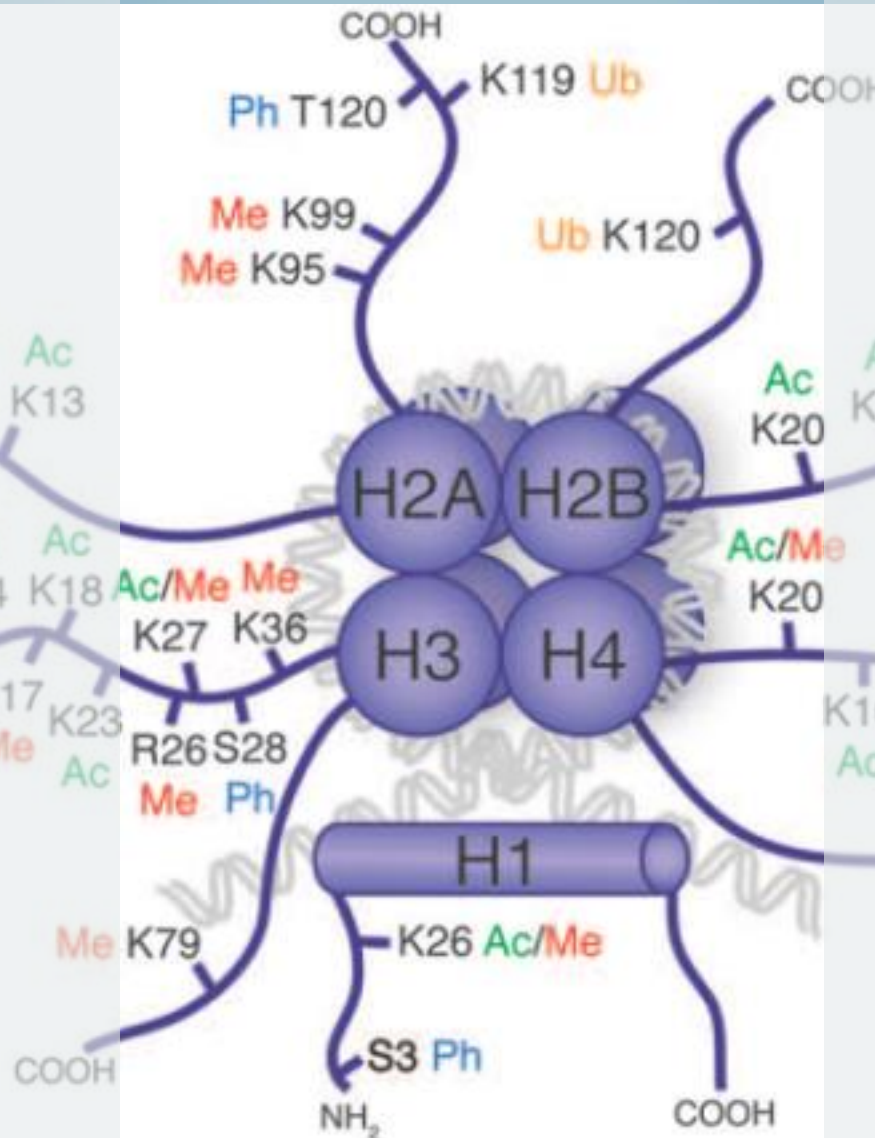
“READERS,” enzymes that recognize, bind and recruit other proteins to the modifications



The recruited proteins then act to alter chromatin structure to promote or repress transcription.

HISTONE MODIFICATIONS

H3-Acetyl-Lys27
 H3-Trimethyl-Lys4
 H3-Monomethyl-Lys27
 H3-Monomethyl-Arg8
 H3-Dimethyl-Lys27
 H3-Dimethyl-Arg8
 H3-Di/Trimethyl-Lys27
 H3-Acetyl-Lys9
 H3-Trimethyl-Lys27
 H3-Panmethyl-Lys9
 H3-Phospho-Ser28
 H3-Monomethyl-Lys9
 H3-Phospho-Ser31
 H3-Dimethyl-Lys9
 H3-Acetyl-Lys36
 H3-Trimethyl-Lys9
 H3-Dimethyl-Lys36
 H3-Phospho-Ser10
 H3-Trimethyl-Lys36
 H3-Phospho-Ser28
 H3-Dimethyl-Arg2
 H3-Monomethyl-Lys18



H3-Monomethyl-Lys79
 H3-Phospho-Thr3
 H3-Acetyl-Lys23
 H3-Dimethyl-Lys79
 H3-Acetyl-Lys4
 H3-Monomethyl-Lys23
 H3-Monomethyl-Lys122
 H3-Monomethyl-Lys4
 H3-Dimethyl-Lys23
 H3-Dimethyl-Lys4
 H3-Phospho-Thr45
 H3-Phospho-Thr11
 H3-Acetyl-Lys56
 H3-Acetyl-Lys14
 H3-Monomethyl-Lys56
 H3-Dimethyl-Lys14
 H3-Dimethyl-Lys56
 H3-Dimethyl-Arg17
 H3-Acetyl-Lys64
 H3-Acetyl-Lys18
 H3-Acetyl-Lys79
 ETC.....

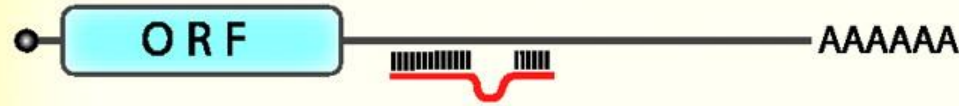
EPIGENETIC MODIFICATION: NON-CODING RNAs

A non-coding RNA is a functional RNA molecule that is not translated into a protein.

siRNAs, microRNAs (~22 nucleotides; fine tune gene expression)

A mechanism for post-transcriptional gene regulation.

imperfect complimentarity = translational repression



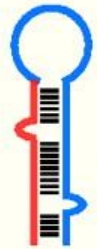
Ago-1



mature
microRNA



Dicer

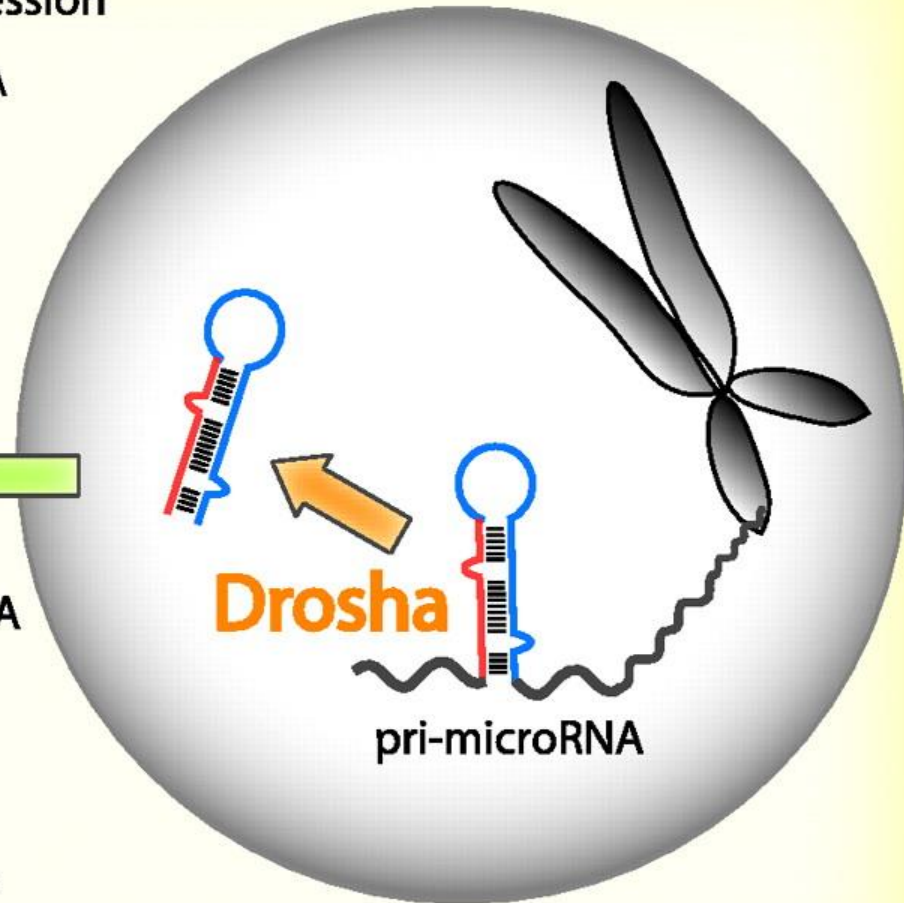


pre-microRNA

Ago-2 (Slicer)



perfect complimentarity = RNA interference



Epigenetics: the missing information?

```
graph TD; A["Epigenetics: the missing information?"] --> B["Lack of identified genetic determinants that fully explain the heritability of complex traits."]; A --> C["Inability to pinpoint causative genetic effects in some complex diseases."]; B --> D["Organisms that face extreme environmental challenges are of particular interest because they survive under conditions incompatible with viability of humans .... and yet there can be only tiny differences in underlying DNA sequences."]; C --> D;
```

Lack of identified genetic determinants that fully explain the heritability of complex traits.

Inability to pinpoint causative genetic effects in some complex diseases.

Organisms that face extreme environmental challenges are of particular interest because they survive under conditions incompatible with viability of humans and yet there can be only tiny differences in underlying DNA sequences.

TURNING OFF GENES: ROLE OF EPIGENETICS

Epigenetics:

- Stable changes in gene activity that do not involve changes in DNA sequence

Common mechanisms:

- DNA methylation
- Histone modification / histone variants
e.g. acetylation, phosphorylation
- Regulatory non-coding RNAs
- “Hiding messages”

Global changes in methylation of gene promoters to reduce transcription rates

Global changes in histone modifications to reduce accessibility to promoter regions by transcription machinery

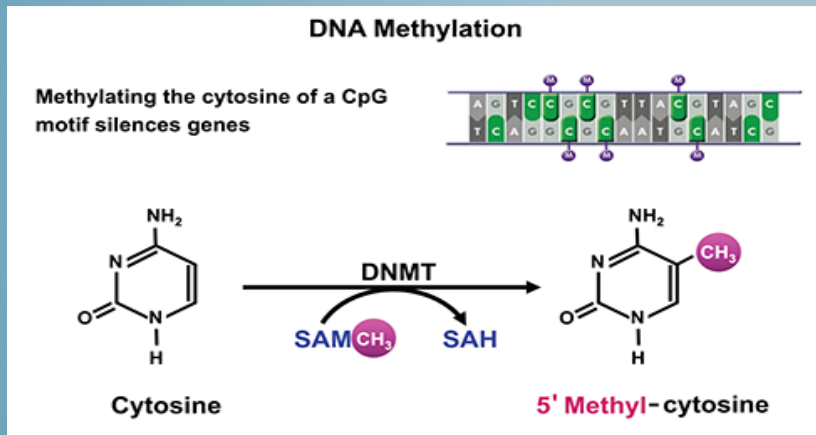
Transcription and translation are ATP-expensive
Epigenetic modifications could alter rates of transcription/translation to produce energy savings in hypometabolism

MicroRNAs can coordinate expression of cell proteins via post-transcriptional action

Other post-transcriptional controls can apply –

- formation of stress granules &
- action of RNA binding proteins

DNA Methylation & Gene Silencing



<http://pubs.niaaa.nih.gov/publications/arcr351/6-16.htm>

Changes in DNA methylation modifies gene transcription

Affects: development, disease, phenotypic plasticity, seasonal changes, behaviour, etc.

Integr Comp Biol, 2014 Jul;54(1):68-76. doi: 10.1093/icb/ictu034. Epub 2014 May 10.

The dynamic nature of DNA methylation: a role in response to social and seasonal variation.

Alvarado S¹, Fernald RD², Storey KB², Szyf M³.

Author information

Abstract

An organism's ability to adapt to its environment depends on its ability to regulate and maintain tissue specific, temporal patterns of gene transcription in response to specific environmental cues. Epigenetic mechanisms are responsible for many of the intricacies of a gene's regulation that alter expression patterns without affecting the genetic sequence. In particular, DNA methylation has been shown to have an important role in regulating early development and in some human diseases. Within these domains, DNA methylation has been extensively characterized over the past 60 years, but the discovery of its role in regulating behavioral outcomes has led to renewed interest in its potential roles in animal behavior and phenotypic plasticity. The conservation of DNA methylation across the animal kingdom suggests a possible role in the plasticity of genomic responses to environmental cues in natural environments. Here, we review the historical context for the study of DNA methylation, its function and mechanisms, and provide examples of gene/environment interactions in response to social and seasonal cues. Finally, we discuss useful tools to interrogate and dissect the function of DNA methylation in non-model organisms.

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Alvarado, S., Fernald, R.D., Storey, K.B., and Szyf, M. 2014.

DNA Methylation & Mammalian Hibernation

[J Exp Biol](#). 2015 Apr 23. pii: jeb.116046. [Epub ahead of print]

Dynamic changes in global and gene specific DNA methylation during hibernation in adult thirteen-lined ground squirrels, *Ictidomys tridecemlineatus*.

Alvarado S¹, Mak T², Liu S², Storey KB³, Szyf M⁴.

Author information

Abstract

Hibernating mammals conserve energy in the winter by undergoing prolonged bouts of torpor, interspersed with brief arousals back to euthermia. These bouts are accompanied with a suite of reversible physiological and biochemical changes; however, much remains to be discovered about the molecular mechanisms involved. Given the seasonal nature of hibernation, it stands to reason that underlying plastic epigenetic mechanisms should exist. One such form of epigenomic regulation involves the reversible modification of cytosine bases in DNA by methylation. DNA methylation is well-known to be a mechanism that confers upon DNA its cellular identity during differentiation in response to innate developmental cues. However, it has recently been hypothesized that DNA methylation also acts as a mechanism for adapting genome function to changing external environmental and experiential signals over different time scales, including during adulthood. Here, we tested the hypothesis that DNA methylation is altered during hibernation in adult wild animals. This study evaluated global changes in DNA methylation in response to hibernation in the liver and skeletal muscle of thirteen-lined ground squirrels along with changes in expression of DNA methyltransferases (DNMT1/3B) and methyl binding domain proteins (MBDs). A reduction in global DNA methylation occurred in muscle during torpor phases whereas significant changes in DNMTs and MBDs were seen in both tissues. We also report dynamic changes in DNA methylation in the promoter of the myocyte enhancer factor 2C (*mef2c*) gene, a candidate regulator of metabolism in skeletal muscle. Taken together, these data show that genomic DNA methylation is dynamic across torpor-arousal bouts during winter hibernation, consistent with a role for this regulatory mechanism in contributing to the hibernation phenotype.

Alvarado, S., Mak, T., Liu, S., Storey, K.B., and Szyf, M. 2015. in press



Changes in
DNA
methylation
& DNMTs
restrict gene
transcription
during torpor

Global changes in methylation of gene promoters to reduce transcription rates

Global changes in histone modifications to reduce accessibility to promoter regions by transcription machinery

Transcription and translation are ATP-expensive
Epigenetic modifications could alter rates of transcription/translation to produce energy savings in hypometabolism

MicroRNAs can coordinate expression of cell proteins via post-transcriptional action

Other post-transcriptional controls can apply

- formation of stress granules &
- action of RNA binding proteins

Histone Deacetylases & Mammalian Hibernation



Available online at www.sciencedirect.com

ScienceDirect

Cryobiology 53 (2006) 310–318

CRYOBIOLOGY

www.elsevier.com/locate/crybio

Evidence for a reduced transcriptional state during hibernation in ground squirrels [☆]

Pier Jr Morin*, Kenneth B. Storey

Institute of Biochemistry and Department of Chemistry, Carleton University, 1125 Colonel By Drive, Ottawa, Ont., Canada K1S 5B6

Received 14 March 2006; accepted 4 August 2006

Available online 18 September 2006

Abstract

During mammalian hibernation, metabolic rate can be reduced to <5% of the euthermic rate as a result of coordinated suppression of multiple energy expensive metabolic processes. Gene transcription is one of these and the present study examines mechanisms of transcriptional control that could contribute to lowering the rate of gene expression in torpor. Histone deacetylases (HDAC) have been linked to gene silencing and measured HDAC activity was 1.82-fold higher in skeletal muscle of hibernating thirteen-lined ground squirrels, *Spermophilus tridecemlineatus*, compared with euthermic controls. Western blotting also showed that HDAC1 and HDAC4 protein levels were 1.21- and 1.48-fold higher, respectively, in muscle from torpid animals. Histone H3 was also evaluated by Western blotting. Total histone H3 was unchanged but two forms of covalently modified histone H3 that are associated with active transcription (phosphorylated Ser 10 and acetylated Lys 23) were significantly reduced by 38–39% in muscle during hibernation. Finally, RNA polymerase II activity was measured using a PCR-based approach; activity in muscle from hibernating squirrels was only 57% of the euthermic value. These data support an overall decrease in transcriptional activity in skeletal muscle of hibernating animals that is accomplished by multiple molecular mechanisms.

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Histone deacetylases allow histones to wrap around DNA more tightly during torpor



Transcription Suppression in Hibernation

- Phospho-Histone H3 (Ser10) levels reduced
 - * **Inhibits transcription**
- Histone Deacetylase activity increased 80%
- Acetyl-Histone H3 (Lys23) levels reduced
 - * **Both inhibit transcription ***
- HDAC 1 & 4 protein levels increased
- RNA Polymerase II activity decreased

Histone Deacetylases & Anoxia Tolerance

Mol Cell Biochem (2010) 342:151–161
DOI 10.1007/s11010-010-0479-5

Epigenetics in anoxia tolerance: a role for histone deacetylases

Anastasia Krivoruchko · Kenneth B. Storey

Histone deacetylases are involved in natural anoxia tolerance

© Springer Science+Business Media, LLC. 2010 / Accepted: 17 April 2010 / Published online: 1 May 2010

Importance of epigenetics has been established in biological processes but the relevance of epigenetic mechanisms to animal survival of low oxygen has not been examined. To establish whether histone deacetylases could be involved in natural anoxia tolerance, we have examined the anoxia-responsive expression of the transcriptional silencers, histone deacetylases (HDACs), in tissues of a unique model for anoxia tolerance,

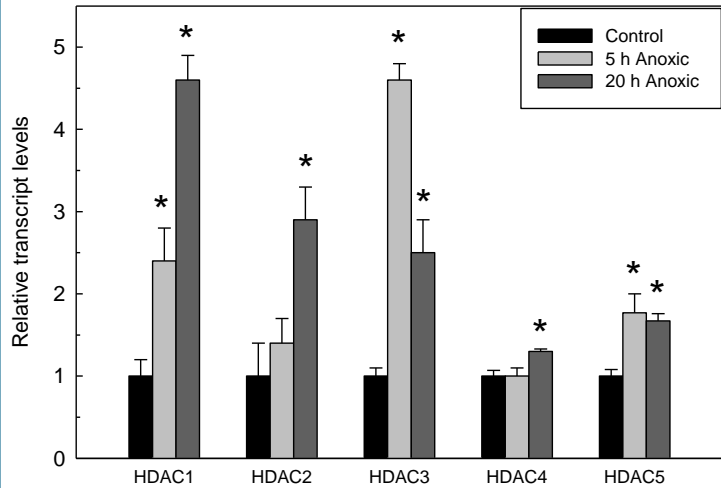
by contributing to this hypometabolic state.

Keywords *Trachemys scripta* · Anoxia tolerance · Epigenetics · Histone deacetylases

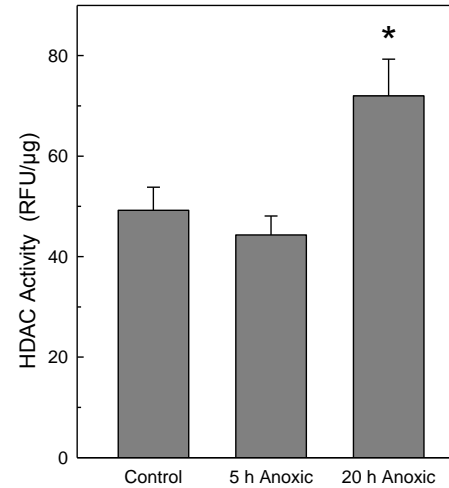


HDAC Responses to anoxia in turtle white muscle

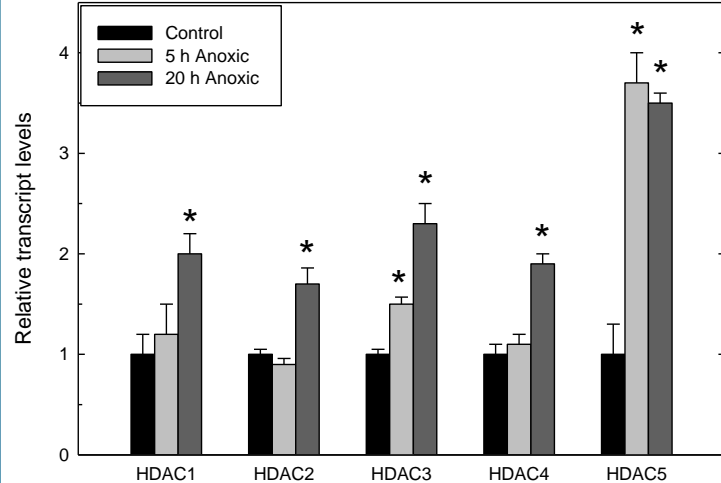
A. MUSCLE HDAC TRANSCRIPT LEVELS



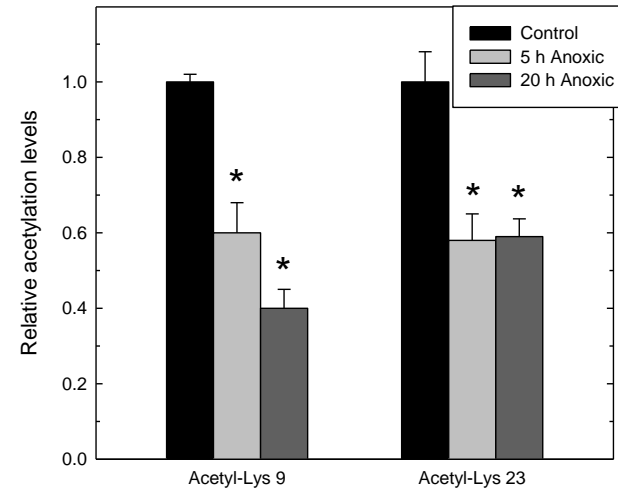
C. MUSCLE HDAC ACTIVITY



B. MUSCLE HDAC PROTEIN LEVELS



D. MUSCLE HISTONE 3 ACETYLATION



Global changes in methylation of gene promoters to reduce transcription rates

Global changes in histone modifications to reduce accessibility to promoter regions by transcription machinery

Transcription and translation are ATP-expensive
Epigenetic modifications could alter rates of transcription/translation to produce energy savings in hypometabolism

MicroRNAs can coordinate expression of cell proteins via post-transcriptional action

Other post-transcriptional controls can apply

- formation of stress granules &
- action of RNA binding proteins

Turning it all off

Journal of Molecular Cell Biology Advance Access published December 21, 2010

doi:10.1093/jmcb/mjq045

Journal of Molecular Cell Biology (2010), 1–9 | 1

Review

The emerging roles of microRNAs in the molecular responses of metabolic rate depression

Kyle K. Biggar and Kenneth B. Storey*

Institute of Biochemistry and Department of Biology, Carleton University, 1125 Colonel By Drive, Ottawa, ON, Canada K1S 5B6

* Correspondence to: Kenneth B. Storey, Tel: +613-520-3678; Fax: +613-520-3749; E-mail: kenneth_storey@carleton.ca

Metabolic rate depression is a key adaptation for survival during hibernation. The molecular mechanisms underlying this process are still poorly understood, but recent studies have shown that metabolic states in hibernating organisms are likely driven by changes in gene expression, particularly in the regulation of metabolism and energy balance. These changes are primarily driven by alterations in the expression of key genes, such as those involved in the cell cycle and DNA damage response. The study of these mechanisms is crucial for understanding the molecular basis of hibernation and its potential applications in medicine and biotechnology.

Metabolic rate depression, a key adaptation for survival during hibernation, is likely driven by changes in gene expression, particularly in the regulation of metabolism and energy balance. These changes are primarily driven by alterations in the expression of key genes, such as those involved in the cell cycle and DNA damage response. The study of these mechanisms is crucial for understanding the molecular basis of hibernation and its potential applications in medicine and biotechnology.

miRNAs & Dicer enzyme show organ-specific changes in mammalian hibernation

Biochimica et Biophysica Acta 1779 (2008) 628–633

Contents lists available at ScienceDirect



Biochimica et Biophysica Acta

journal homepage: www.elsevier.com/locate/bbagrm



Differential expression of microRNA species in organs of hibernating ground squirrels: A role in translational suppression during torpor

Pier Jr. Morin, Adrian Dubuc, Kenneth B. Storey*

Institute of Biochemistry and Department of Chemistry, Carleton University, 1125 Colonel By Drive, Ottawa, Ontario, Canada K1S 5B6

ARTICLE INFO

Article history:

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Available online 5 August 2008

Keywords:

MicroRNA

Hibernation

Spermophilus tridecemlineatus

Dicer

Reversible control of translation

ABSTRACT

Mammalian hibernation includes long periods of profound torpor where the rates of all metabolic processes are strongly suppressed in a reversible manner. We hypothesized that microRNAs (miRNAs), small non-coding transcripts that bind to mRNA, could play a role in the global suppression of mRNA translation when animals enter torpor. Selected miRNA species (4–9 of the following: mir-1, mir-24, mir-15a, mir-16, mir-21, mir-122a, mir-143, mir-146 and mir-206) were evaluated in four organs of euthermic versus hibernating ground squirrels, *Spermophilus tridecemlineatus* using RT-PCR. Levels of mir-24 transcripts were significantly reduced in heart and skeletal muscle of torpid animals as were mir-122a levels in the muscle. Mir-1 and mir-21 both increased significantly in kidney during torpor by 2.0- and 1.3-fold, respectively. No changes were found for the four miRNA species analyzed in liver. Protein levels of Dicer, an enzyme involved in miRNA processing were also quantified in heart, kidney and liver. Dicer protein levels increased by 2.7-fold in heart during hibernation but decreased by 60% in kidney. These data are the first report that differential regulation



ARE MicroRNAs DIFFERENTIALLY REGULATED IN HIBERNATORS?

- **Yes!** Selected miRNAs were regulated in heart, muscle & kidney of hibernating 13-lined ground squirrels

miRNA	Fold change	Process in higher vertebrates
Mir-1	2.0	Myogenesis
Mir-133a	2.4	Myogenesis
Mir-206	2.6	Myogenesis
Let-7	2.0	Cell cycle
Mir-26	2.4	Hypoxia
Mir-451	2.6	Erythropoiesis

(Morin, Dubuc & Storey, 2008, Biochim Biophys Acta 1779:628-633)

MicroRNAs and Regulation of the Cell Cycle

Current Genomics, 2009, 10, 573-584

573

Perspectives in Cell Cycle Regulation: Lessons from an Anoxic Vertebrate

Kyle K. Biggar and Kenneth B. Storey*

Institute of Biochemistry and Department of Biology, Carleton University, 1125 Colonel By Drive, Ottawa, ON, K1S 5B6, Canada

Abstract: The ability of an animal, normally dependent on aerobic respiration, to suspend breathing and enter an anoxic state for long term survival is clearly a fascinating feat, and has been the focus of numerous biochemical studies. When anoxia tolerant turtles are faced with periods of oxygen deprivation, numerous physiological and biochemical alterations take place in order to facilitate vital reductions in ATP consumption. Such strategies include reversible post-translational modifications as well as the implementation of translation and transcription controls facilitating metabolic depression. Although it is clear that anoxic survival relies on the suppression of ATP consuming processes, the state of the cell cycle in anoxia tolerant vertebrates remain elusive. Several anoxia tolerant invertebrate and embryonic vertebrate models display cell cycle arrest when presented with anoxic stress. Despite this, the cell cycle has not yet been characterized for anoxia tolerant vertebrates. This review article will discuss the possibility of translational control of the cell cycle in anoxia tolerant vertebrate tissues. Consequentially, the possibility of translational control of the cell cycle in anoxia tolerant vertebrate tissues. Consequentially, the possibility of translational control of the cell cycle in anoxia tolerant vertebrate tissues. Consequentially, the possibility of translational control of the cell cycle in anoxia tolerant vertebrate tissues.



Cell Cycle 11:9, 1705-1713; May 1, 2012; © 2012 Landes Bioscience

SPECIAL FOCUS REPORT

Evidence for cell cycle suppression and microRNA regulation of cyclin D1 during anoxia exposure in turtles

Kyle K. Biggar and Kenneth B. Storey*

Institute of Biochemistry and Department of Biology, Carleton University, Ottawa, ON Canada

Key words: *Trachemys scripta elegans*, anoxia, microRNA, cyclin, metabolic rate depression

The red-eared slider turtle (*Trachemys scripta elegans*) has a well-developed natural tolerance for oxygen deprivation that derives from biochemical adaptations, including anoxia-induced suppression of metabolic rate. We hypothesized that mechanisms that suppress ATP-expensive cell cycle activity would contribute significantly to establishing the hypometabolic state during anaerobiosis. Cyclin D1 is a critical regulator of the G₁ phase of the cell cycle and is regarded as key to initiating cell proliferation. The relative protein expression of cyclin D1 was analyzed in both whole-cell and nuclear fractions of liver, kidney and skeletal muscle from turtles exposed to 5 or 20 h of submergence anoxia. Expression of cyclin D1 in both total and nuclear fractions decreased significantly under anoxia in liver and kidney as compared to normoxic controls. The relative phosphorylation state of cyclin D1 in muscle. The relative phosphorylation state of cyclin D1 in muscle. The relative phosphorylation state of cyclin D1 in muscle. Since phosphorylation of threonine 286 is necessary for alternative mechanism is responsible for cyclin D1 suppression change under anoxia in any tissue, so a post-transcriptional regulation of cyclin D1 showed the presence of both an AU-rich region and a 5' UTR. Levels of both microRNAs increased in liver and kidney of anoxic turtle. microRNA inhibition of mRNA translation as the mechanism of cyclin D1 suppression in anoxic turtle.

Anoxia elevated miR-16-1 & miR-15a to suppress cyclin D1 protein, a key regulator of cell cycle initiation

MicroRNAs and estivation



Gene 529 (2013) 269–275

Contents lists available at ScienceDirect

Gene

journal homepage: www.elsevier.com/locate/gene

Dehydration mediated microRNA response in the African clawed frog *Xenopus laevis*

Cheng-Wei Wu, Kyle K. Biggar, Kenneth B. Storey *

Institute of Biochemistry and Department of Biology, Carleton University, 1125 Colonel By Drive, Ottawa, ON K1S 5B6, Canada

ARTICLE INFO

Article history:
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Keywords:
Metabolic rate depression
Amphibian
Luminescence
Non-coding RNA

ABSTRACT

Exposure to various environmental stresses induces metabolic rate depression in many animal species, an adaptation that conserves energy until the environment is again conducive to normal life. The African clawed frog *Xenopus laevis*, is periodically subjected to arid summers in South Africa, and utilizes a hypometabolic state of estivation as a mechanism of long term survival. During estivation, frogs must deal with substantial dehydration as their ponds dry out and *X. laevis* can endure >30% loss of its body water. We hypothesize that microRNAs play a vital role in establishing a reversible hypometabolic state and to dehydration stress that is associated with amphibian estivation. The present study analyzes the whole body dehydration on microRNA expression in three tissues of *X. laevis*. Compared to control, miR-1, miR-125b, and miR-16-1 decreased to 37 ± 6 , 64 ± 8 , and $80 \pm 4\%$ of control levels during dehydration in liver. By contrast, miR-210, miR-34a and miR-21 were significantly elevated by 3.05 ± 0.45 , $2.17 \pm 1.36 \pm 0.05$ -fold, respectively, in the liver. In kidney tissue, miR-29b, miR-21, and miR-203 were 1.40 ± 0.09 , 1.31 ± 0.05 , and 2.17 ± 0.31 -fold, respectively, in response to dehydration whereas miR-34a were elevated in ventral skin by 1.35 ± 0.05 and 1.74 ± 0.12 -fold, respectively. Bioinformatics analysis of the differentially expressed microRNAs suggests that these are mainly involved in two processes: (1) regulation of solute carrier proteins, and (2) regulation of mitogen-activated protein kinase signaling. This is the first report that shows a tissue specific mode of microRNA expression during amphibian dehydration. The present study provides evidence for microRNAs as crucial regulators of metabolic depression.

Dehydration led to differential expression of microRNAs in *X. laevis* organs

MicroRNAs and freeze tolerance

Cryobiology 59 (2009) 317–321

Contents lists available at ScienceDirect

Cryobiology

journal homepage: www.elsevier.com/locate/ycryo

MicroRNA regulation below zero: Differential expression of miRNA-21 and miRNA-16 during freezing in wood frogs[☆]

Kyle K. Biggar, Adrian Dubuc, Kenneth Storey*

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ARTICLE INFO

Article history:
Received 1 July 2009
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Available online 6 September 2009

Keywords:
Rana sylvatica
Vertebrate freeze tolerance
MicroRNA
Post-transcriptional control of gene expression
Metabolic rate depression
Anti-apoptosis
Cell cycle control
Dicer

ABSTRACT

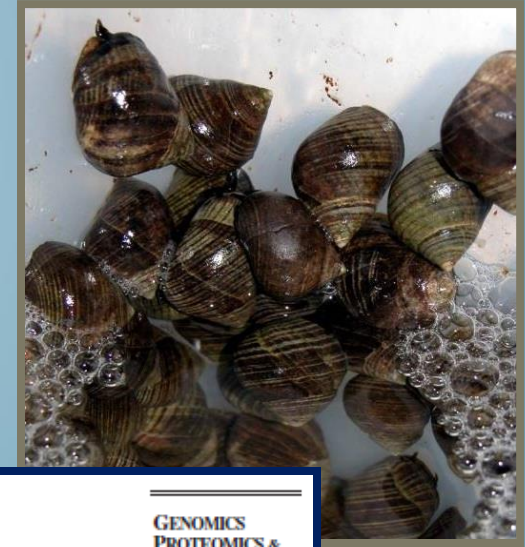
Natural freeze tolerance depends on numerous biochemical adaptations that address the multiple stresses imposed on cells by freezing and preserves viability by suppressing energy-expensive cell functions during the frozen state. We hypothesized that microRNAs, small non-coding RNA transcripts that bind to messenger RNA to act to establish rapid biological controls that aid the reorganization of metabolic priorities for freeze survival. Selected microRNA species (miR-16 and miR-21) were evaluated using RT-PCR in liver and skeletal muscle of wood frogs (*Rana sylvatica*) comparing controls (5 °C acclimated) with animals frozen for 24 h. Levels of miR-21 increased significantly during freezing in both tissues, while levels of miR-16 increased in skeletal muscle but not in liver. miR-16 transcripts also rose significantly in skeletal muscle. Protein levels of Dicer, a type III RNase III in the cytoplasm, were unchanged in liver and decreased in skeletal muscle, representing the first report of differential regulation of microRNAs during freezing. This study provides a mechanism for rapid, yet reversible, gene silencing during freezing.

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miRNAs & Dicer enzyme show organ-specific changes in freeze tolerant frogs



Invertebrate microRNAs: new method for detection & amplification



Analytical Biochemistry 416 (2011) 231–233

Contents lists available at ScienceDirect

Analytical Biochemistry

journal homepage: www.elsevier.com/locate/yabio



Notes & Tips

Amplification and sequencing of mature microRNAs in uncharacterized animal models using stem–loop reverse transcription–polymerase chain reaction

Kyle K. Biggar, Samantha F. Kornfeld, Kenneth B. Storey*

Institute of Biochemistry and Department of Biology, Carleton University, Ottawa, Ontario, Canada K1S 5B6

ARTICLE INFO

Article history:
Received 8 April 2011
Received in revised form 10 May 2011

Accepted

Available

ABSTRACT

Expression of mature microRNA (miRNA) in animal model systems but is difficult to evaluate using conventional methods. We have developed a stem–loop reverse transcription–polymerase chain reaction (SL-RT-PCR) method for the detection and amplification of mature miRNAs in uncharacterized animal models. This method provides a rapid and sensitive method for the detection and amplification of mature miRNAs in uncharacterized animal models.

MicroRNAs
respond to
anoxia &
freezing in
intertidal snails



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Genomics Proteomics Bioinformatics 10 (2012) 302–309

GENOMICS
PROTEOMICS &
BIOINFORMATICS

www.elsevier.com/locate/gpb

Original Research

MicroRNA Regulation in Extreme Environments:
Differential Expression of MicroRNAs in the Intertidal Snail *Littorina littorea* During Extended Periods of Freezing and Anoxia

Kyle K. Biggar^{#,*}, Samantha F. Kornfeld[#], Yulia Maistrovski, Kenneth B. Storey

Institute of Biochemistry & Department of Biology, Carleton University, Ottawa, ON K1S 5B6, Canada

Received 26 July 2012; revised 11 September 2012; accepted 12 September 2012

Available online 8 October 2012

Abstract

Several recent studies of vertebrate adaptation to environmental stress have suggested roles for microRNAs (miRNAs) in regulating global suppression of protein synthesis and/or restructuring protein expression patterns. The present study is the first to characterize stress-responsive alterations in the expression of miRNAs during natural freezing or anoxia exposures in an invertebrate species, the intertidal gastropod *Littorina littorea*. These snails are exposed to anoxia and freezing conditions as their environment constantly fluctuates on both a tidal and seasonal basis. The expression of selected miRNAs that are known to influence the cell cycle, cellular signaling pathways, and metabolism was determined using RT-PCR. Compared to control, significant changes in expression were observed in several miRNAs during both freezing and anoxia exposures.

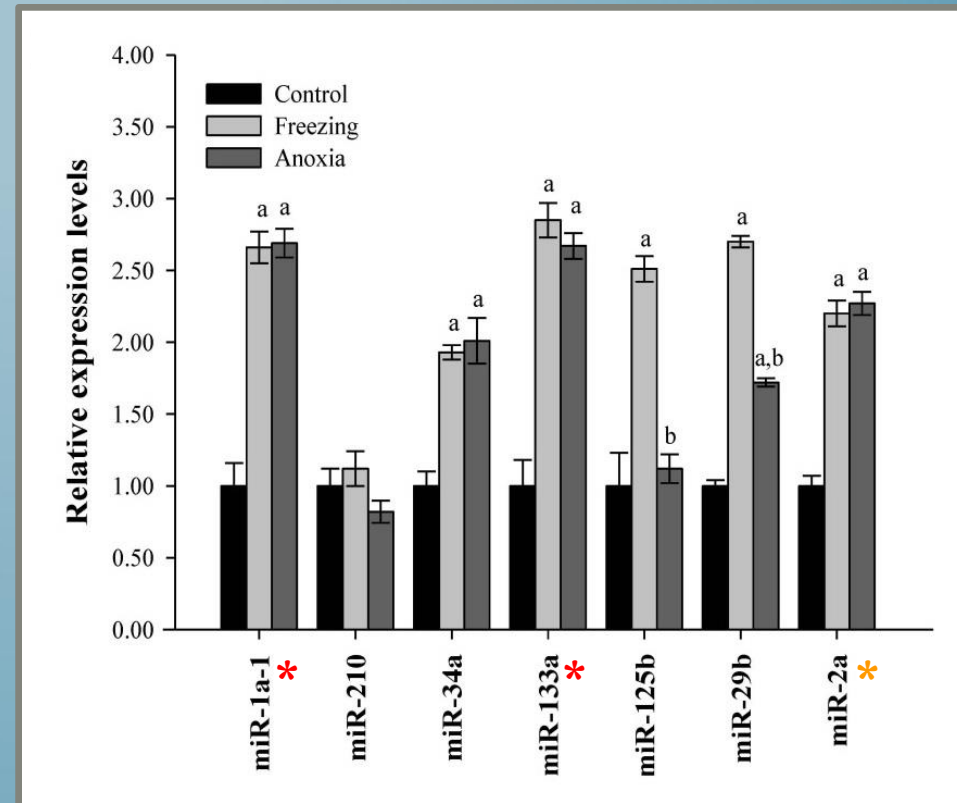
MicroRNAs in *Littorina littorea* FOOT MUSCLE: Up-regulated by Freezing & Anoxia

miR-1a-1* & miR-133a*

- myocyte proliferation & differentiation
- regulate *Mef2a* and *Gata4*, Tfs that promote muscle maintenance

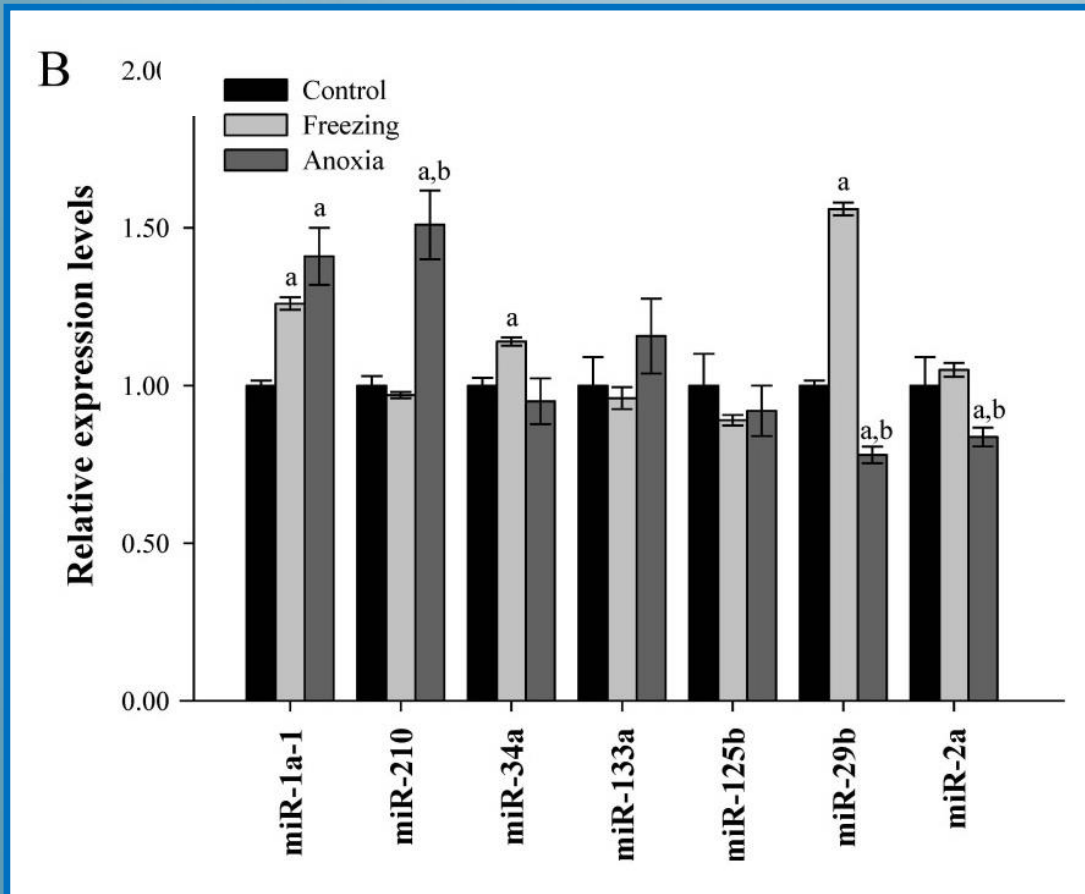
miR-2a*

- anti-apoptotic action by targeting the pro-apoptotic protein, *Reaper*



Biggar, Kornfeld & Storey, 2011. Anal. Biochem. 416, 231-3.
Biggar, Kornfeld, Maistrovski & Storey, 2012. Genom.
Proteom. Biotech. in press

Micro RNAs in *Littorina littorea* HEPATOPANCREAS: Up-regulated by Freezing & Anoxia

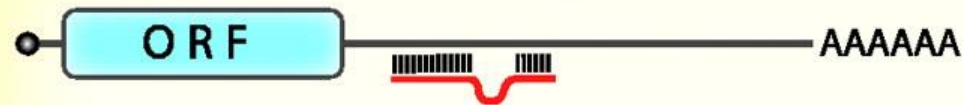


Major changes:

- miR-1a-1 up in freeze & anoxia (like in foot)
- miR-210 up in anoxia
- miR-29b up in freeze

MICRO RNA: Drosha & Dicer

imperfect complimentarity = translational repression



Ago-1 ↑

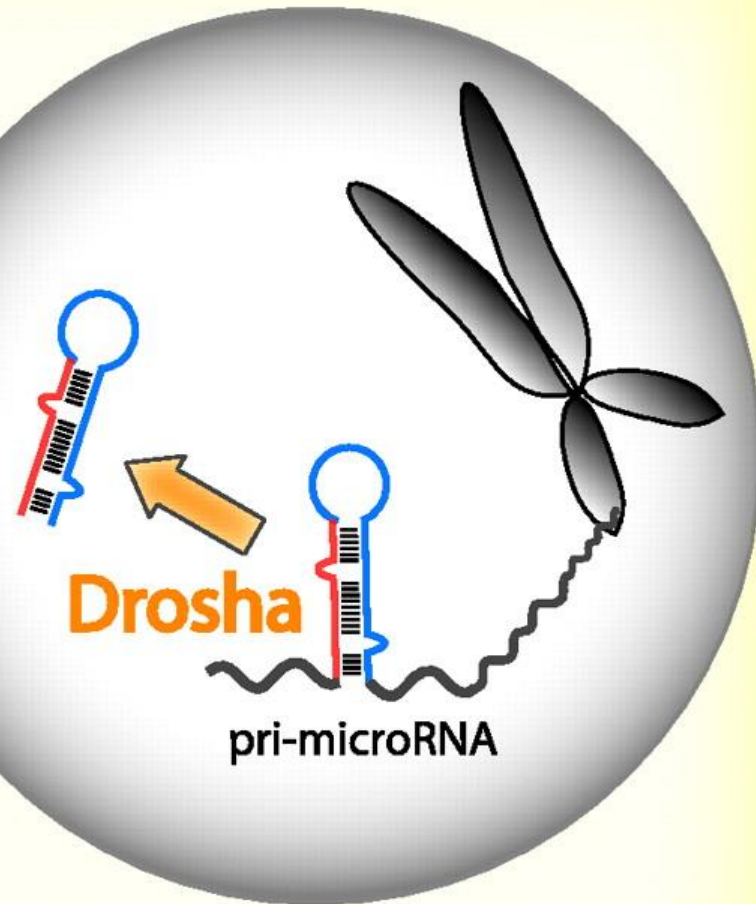
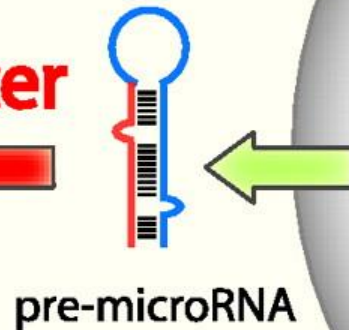
mature microRNA

Ago-2 (Slicer) ↓

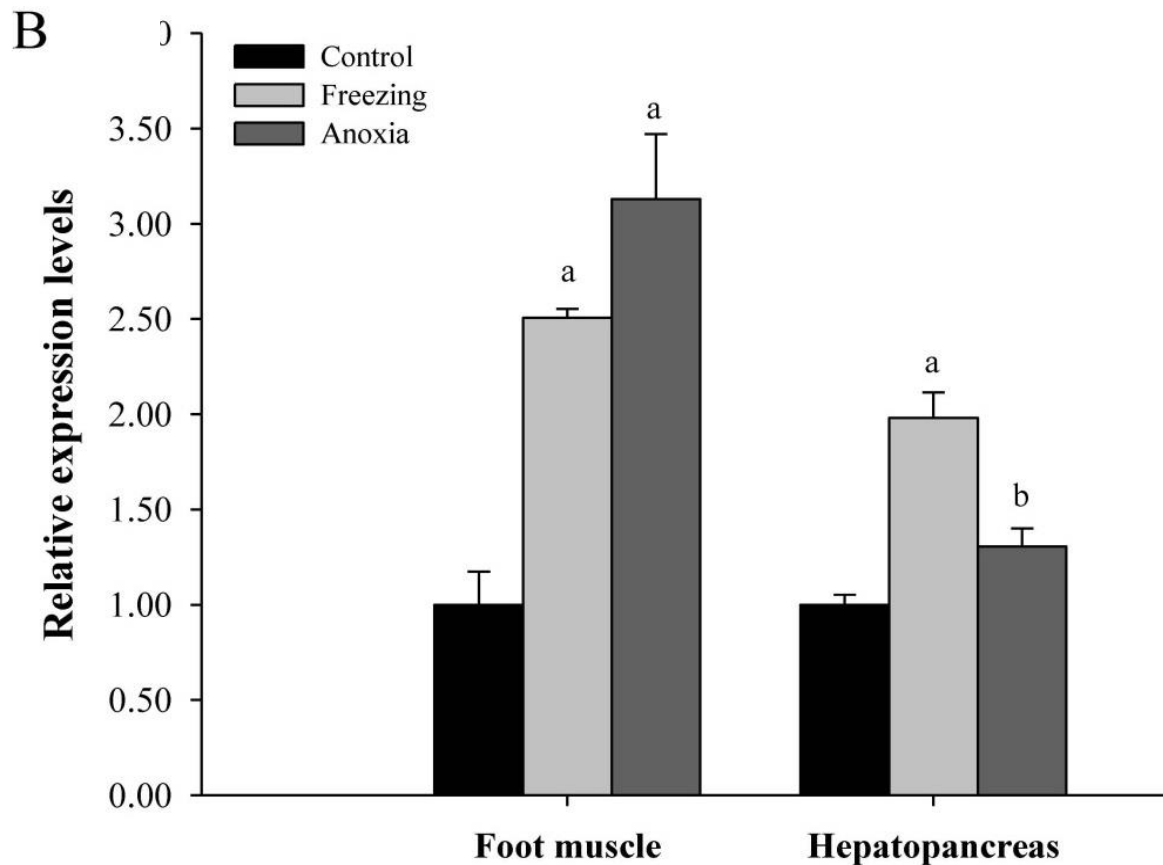


perfect complimentarity = RNA interference

Dicer ←



DICER ENZYME IN *L. littorea* TISSUES



Dicer protein
increased in
both freezing
& anoxia
(immunoblots)

Elevated miRNA
processing

MicroRNAs and aestivation



Marine Genomics xxx (2014) xxx–xxx

Contents lists available at ScienceDirect

Marine Genomics

journal homepage: www.elsevier.com/locate/margen

Short communication

Large-scale identification and comparative analysis of miRNA expression profile in the respiratory tree of the sea cucumber *Apostichopus japonicus* during aestivation

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^a Fisheries College, Ocean University of China, Qingdao, PR China
^b Institute of Biochemistry, Carleton University, 1125 Colonel By Drive, Ottawa, ON K1S 5B6, Canada

ARTICLE INFO

ABSTRACT

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Available online xxxc

Keywords:
miRNA
Sea cucumber
Aestivation

The sea cucumber *Apostichopus japonicus* withstands high metabolic rate and entering a state of aestivation. We hypothesize that this process involves important post-transcriptional regulation of over mRNA translation. The present study analyzed profiles of miRNAs specific to sea cucumber. Animals sampled during aestivation (torpor) were compared with animals from a non-aestivation (active) state. We identified 308 sea cucumber miRNAs, including 18 novel miRNAs specific to sea cucumber. Among the most prominent miRNA species, miR-200-3p, miR-2004, miR-2010, miR-22, miR-252a, miR-252a-3p and miR-92 were significantly over-expressed during deep aestivation compared with non-aestivation animals. Preliminary analyses of their putative target genes and GO analysis suggest that these miRNAs could play important roles in global transcriptional depression and cell differentiation during aestivation. High-throughput sequencing data and microarray data have been submitted to GEO database.

OPEN ACCESS Freely available online

PLOS ONE

High-Throughput Sequencing Reveals Differential Expression of miRNAs in Intestine from Sea Cucumber during Aestivation

Muyan Chen^{1*}, Xiumei Zhang¹, Jianning Liu², Kenneth B. Storey³

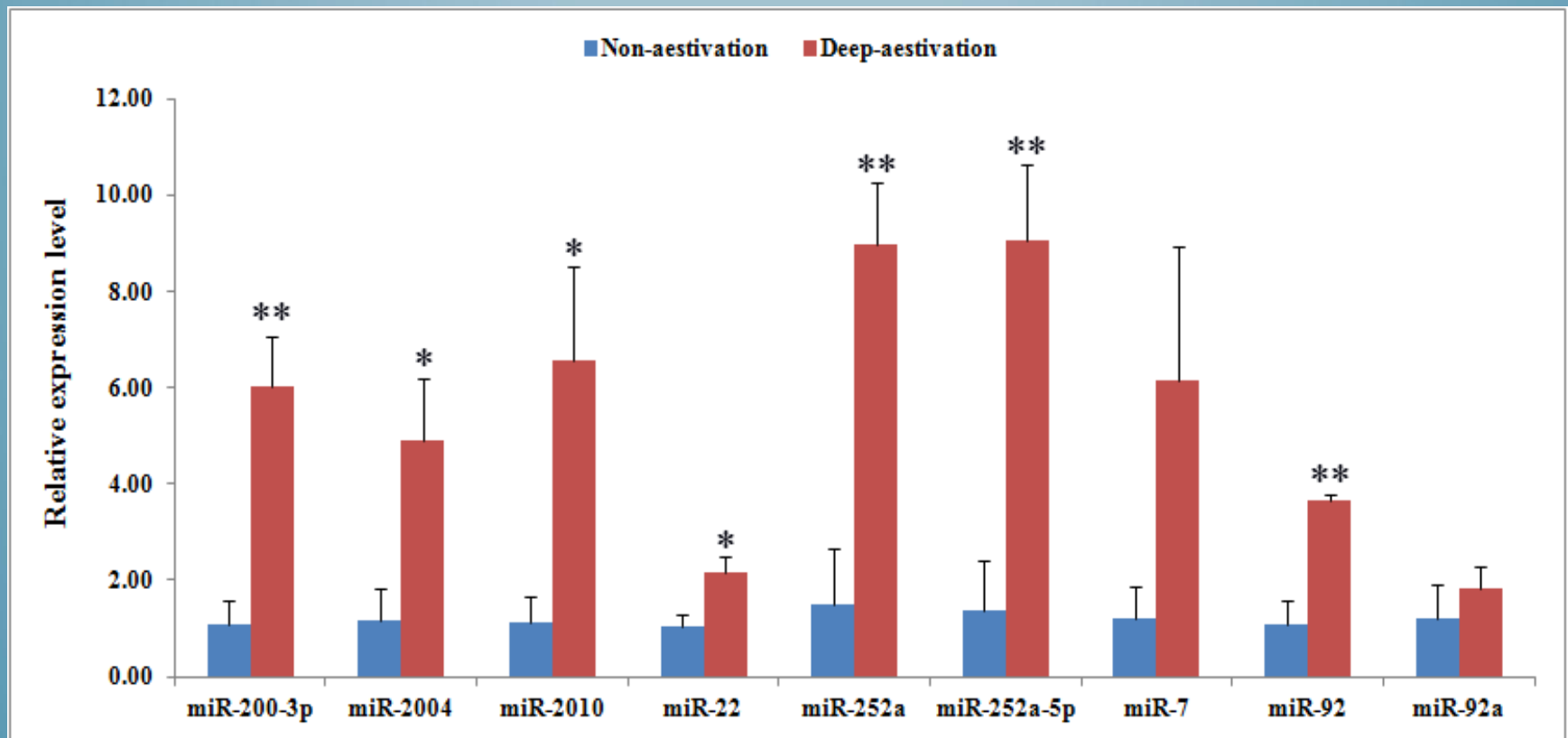
¹ Fisheries College, Ocean University of China, Qingdao, PR China, ² LC-BIO CO., LTD. Hangzhou, PR China, ³ Institute of Biochemistry, Carleton University, Ottawa, Ontario, Canada

Abstract

The regulatory role of miRNA in gene expression is an emerging hot new topic in the control of hypometabolism. Sea cucumber aestivation is a complicated physiological process that includes obvious hypometabolism as evidenced by a decrease in the rates of oxygen consumption and ammonia nitrogen excretion, as well as a serious degeneration of the intestine into a very tiny filament. To determine whether miRNAs play regulatory roles in this process, the present study analyzed profiles of miRNA expression in the intestine of the sea cucumber (*Apostichopus japonicus*), using Solexa deep sequencing technology. We identified 308 sea cucumber miRNAs, including 18 novel miRNAs specific to sea cucumber. Animals sampled during deep aestivation (DA) after at least 15 days of continuous torpor, were compared with animals from a non-aestivation (NA) state (animals that had passed through aestivation and returned to the active state). We identified 42 differentially expressed miRNAs [RPM (reads per million) > 10, |FC| (fold change) ≥ 1, FDR (false discovery rate) < 0.01] during aestivation, which were validated by two other miRNA profiling methods: miRNA microarray and real-time PCR. Among the most prominent miRNA species, miR-200-3p, miR-2004, miR-2010, miR-22, miR-252a, miR-252a-3p and miR-92 were significantly over-expressed during deep aestivation compared with non-aestivation animals. Preliminary analyses of their putative target genes and GO analysis suggest that these miRNAs could play important roles in global transcriptional depression and cell differentiation during aestivation. High-throughput sequencing data and microarray data have been submitted to GEO database.

Tissue specific over-expression of selected microRNAs in aestivation

Aestivation in sea cucumbers: Differential expression microRNAs in intestine



Histone Modification & Hypoxia in Ocean Squid

Reduced phosphorylation or acetylation of histone tails to suppress gene expression under hypoxia

Metabolic suppression during protracted exposure to hypoxia in the jumbo squid, *Dosidicus gigas*, living in an oxygen minimum zone

Brad A. Seibel^{1,*}, N. Sören Häfker², Katja Trübenbach³, Jing Zhang⁴, Shannon N. Tessier⁴, Hans-Otto Pörtner², Rui Rosa³ and Kenneth B. Storey⁴

ABSTRACT

The jumbo squid, *Dosidicus gigas*, can survive extended forays into the oxygen minimum zone (OMZ) of the Eastern Pacific Ocean. Previous studies have demonstrated reduced oxygen consumption and a limited anaerobic contribution to ATP production, suggesting the capacity for substantial metabolic suppression during hypoxic exposure. Here, we provide a more complete description of energy metabolism and explore the expression of proteins indicative of transcriptional and translational arrest that may contribute to metabolic suppression. We demonstrate a suppression of total ATP demand under hypoxic conditions (1% oxygen, $P_{O_2}=0.8$ kPa) in both juveniles (52%) and adults (35%) of the jumbo squid. Oxygen consumption rates are reduced to 20% under hypoxia relative to air-saturated controls. Concentrations of arginine phosphate (Arg-P) and ATP declined initially, reaching a new steady state (~30% of controls) after the first hour of hypoxic exposure. Octopine began accumulating after the first hour of hypoxic exposure, once Arg-P breakdown resulted in sufficient free arginine for substrate. Octopine reached levels near 30 mmol g⁻¹ after 3.4 h of hypoxic

J. Exp. Biol. 217:2555-68; 2014



photo: Scott Cassell

Global changes in methylation of gene promoters to reduce transcription rates

Global changes in histone modifications to reduce accessibility to promoter regions by transcription machinery

Transcription and translation are ATP-expensive
Epigenetic modifications could alter rates of transcription/translation to produce energy savings in hypometabolism

MicroRNAs can coordinate expression of cell proteins via post-transcriptional action

Other post-transcriptional controls can apply

- **formation of stress granules &**
- **action of RNA binding proteins**

Non-coding RNA: MicroRNA & Antisense RNA regulate HIF-1 α in hibernation

J Comp Physiol B. 2012 Aug;182(6):849-59. doi: 10.1007/s00360-012-0662-y. Epub 2012 Apr 13.

HIF-1 α regulation in mammalian hibernators: role of non-coding RNA in HIF-1 α control during torpor in ground squirrels and bats.

Maistrovski Y¹, Biggar KK, Storey KB.

Author information

Abstract

A potential role for non-coding RNAs, miR-106b and antisense hypoxia inducible transcription factor-1 (HIF-1 α), in HIF-1 α regulation during mammalian hibernation was investigated in two species, the thirteen-lined ground squirrel (*Ictidomys tridecemlineatus*) and the little brown bat (*Myotis lucifugus*). Both species showed differential regulation of HIF-1 α during hibernation. HIF-1 α protein levels increased significantly in skeletal muscle of both species when animals entered torpor, as well as in bat liver. HIF-1 α mRNA levels correlated with the protein increase in bat skeletal muscle and liver but not in squirrel skeletal muscle. Antisense HIF-1 α transcripts were identified in skeletal muscle of both hibernators. The expression of



Polysome profiles and mammalian hibernation

[Arch Biochem Biophys.](#) 2002 May 15;401(2):244-54.

The translation state of differentially expressed mRNAs in the hibernating 13-lined ground squirrel (*Spermophilus tridecemlineatus*).

[Hittel D¹](#), [Storey KB](#).

Author information 

Abstract

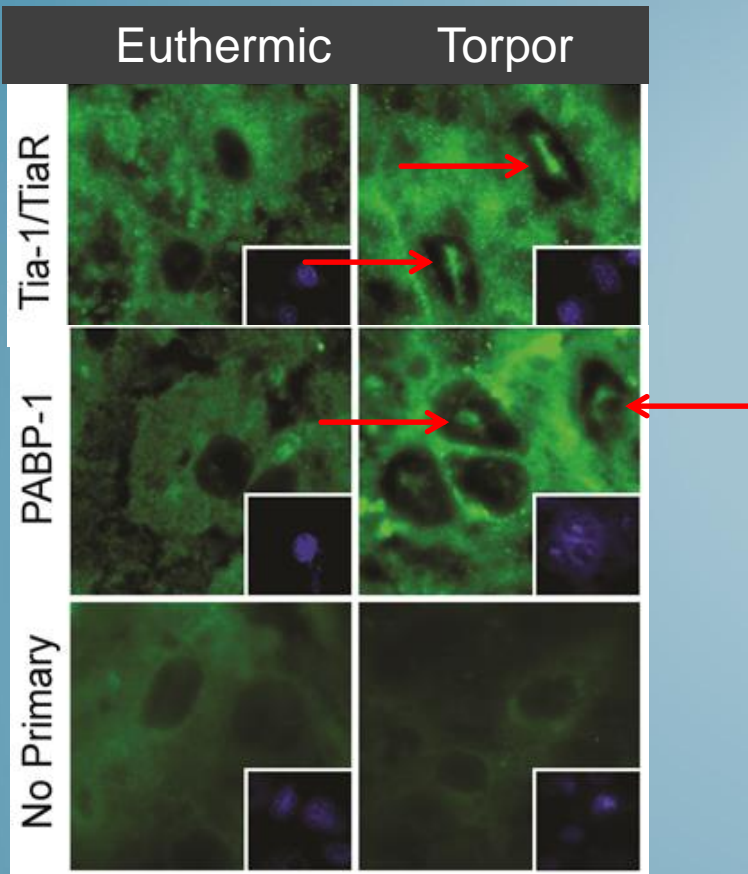
The translation state of differentially expressed mRNAs were compared in kidney and brown adipose tissue of the hibernating ground squirrel, *Spermophilus tridecemlineatus*. Polysome analysis revealed a striking disaggregation of polyribosomes during hibernation and the redistribution of Cox4 (cytochrome c oxidase subunit 4) and Oct2 (organic cation transporter type 2) transcripts into monosome and mRNP fractions of kidney cytoplasmic extracts. Additionally, OCT2 protein levels decreased in kidney of hibernating animals in a rate compared with euthermic kidney. There was no translational depression in brown adipose tissue. H-FABP (fatty acid-binding protein (H-FABP), that is up-regulated during hibernation, was increasingly abundant in the brown adipose tissue. The existence of a tissue-specific mechanism for the regulation of protein synthesis during hibernation.

Polysomes dissociate & mRNA moves to monosome & RNP fractions during torpor



Brown adipose retains polysomes & translation of key proteins e.g. FABP

RNA binding proteins & hibernation



Subnuclear structures formed with TIA & PABP greatly increased during torpor

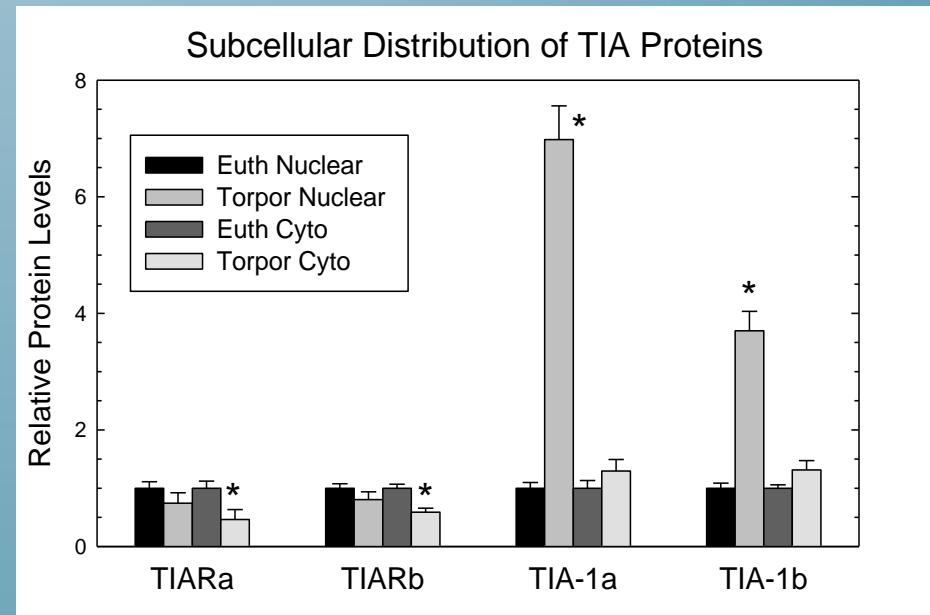
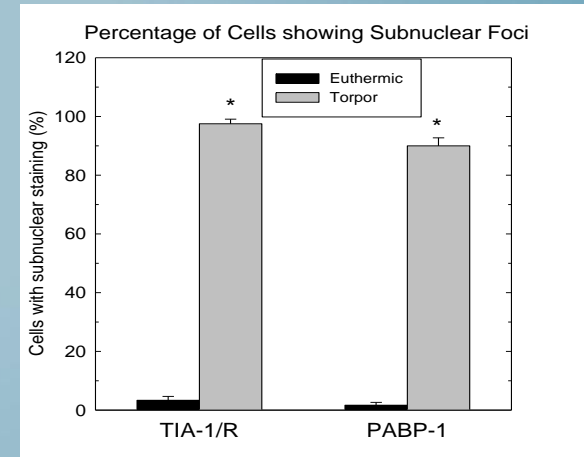
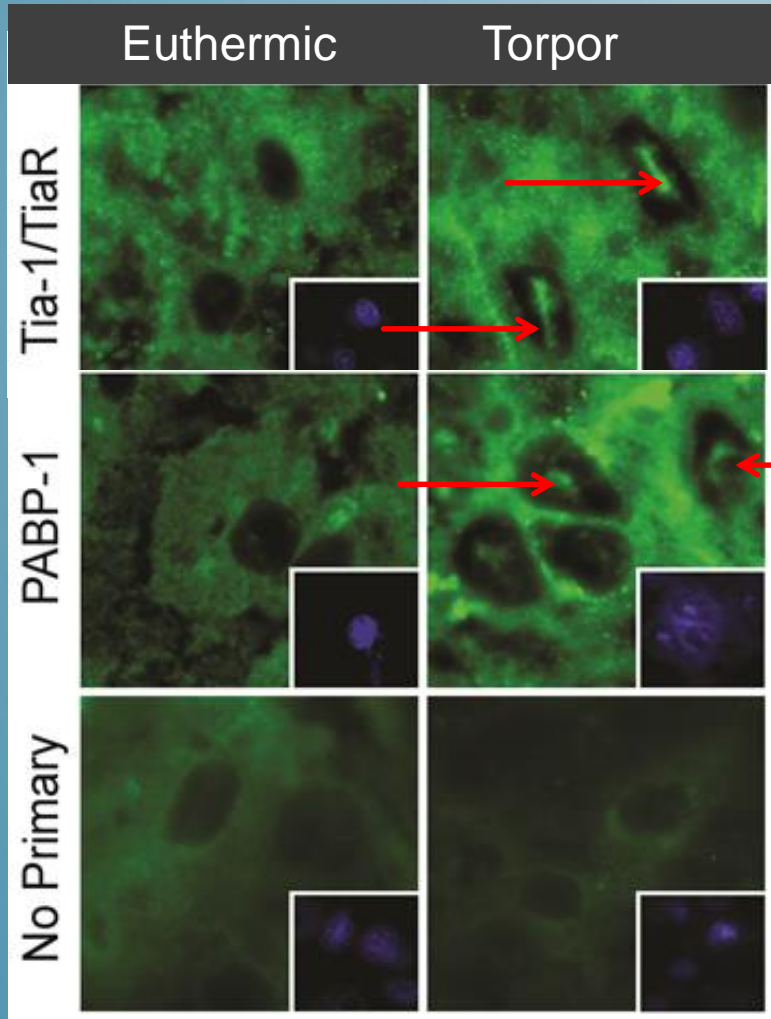
Cell Stress and Chaperones 2014; 19(6):813-25.

ORIGINAL PAPER

The involvement of mRNA processing factors TIA-1, TIAR, and PABP-1 during mammalian hibernation

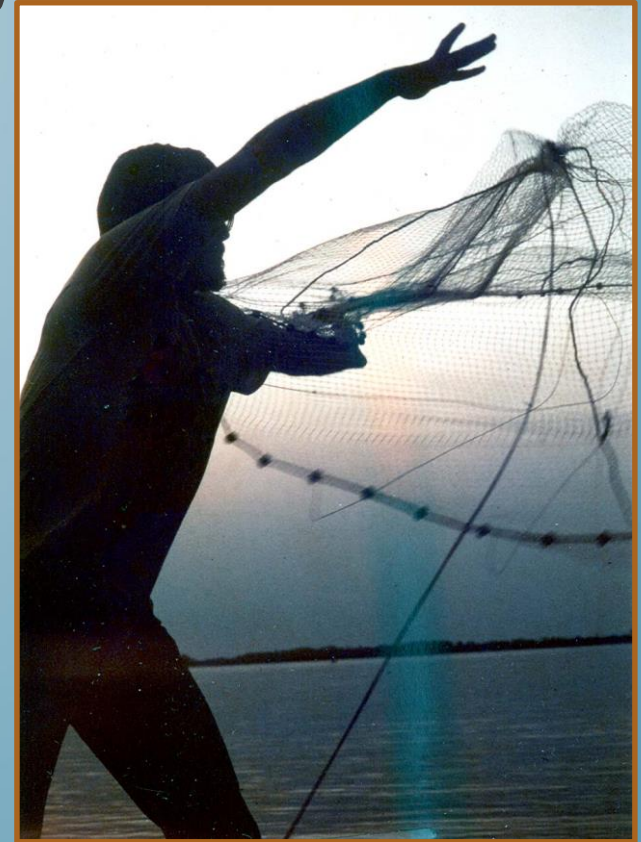
Shannon N. Tessier • Timothy E. Audas • Cheng-Wei Wu •
Stephen Lee • Kenneth B. Storey

RNA Binding Proteins & Mammalian Hibernation



WHERE DO WE GO FROM HERE?

- *Applications of MRD research*
- *Novel phosphorylations*
- *Atrophy, hypertrophy*
-- *autophagy for survival*
- *Turning it all off -- microRNA*
- *Epigenetics & adaptation*
- *Life span extension*
- *Antioxidant Defense*
- *Cell cycle suppression*
- *Unity through evolution*



NEW DIRECTIONS

NEW DIRECTIONS - TBA

Big Science Edition:

1. GENOMES: Sequence all the genes to feel better !

The western painted turtle genome, a model for the evolution of extreme physiological adaptations in a slowly evolving lineage.

***Shaffer HB, ...Storey KB, et al., Genome Biol. 2013
14(3): R28***

2. Protein 2D: What about the Proteins

OMICS – Proteomics

Genomic tools to discover biochemical adaptations

[Genome Biol.](#) 2013 Mar 28;14(3):R28. [Epub ahead of print]

The western painted turtle genome, a model for the evolution of extreme physiological adaptations in a slowly evolving lineage.

[Shaffer HB](#)¹, [Minx P](#), [Warren DE](#), [Shedlock AM](#), [Thomson RC](#), [Valenzuela N](#), [Abramyan J](#), [Amemiya CT](#), [Badenhorst D](#), [Biggar KK](#), [Borchert GM](#), [Botka CW](#), [Bowden RM](#), [Braun EL](#), [Bronikowski AM](#), [Bruneau BG](#), [Buck LT](#), [Capel B](#), [Castoe TA](#), [Czerwinski M](#), [Delehaunty KD](#), [Edwards SV](#), [Fronick CC](#), [Fujita MK](#), [Fulton L](#), [Graves TA](#), [Green RE](#), [Haerty W](#), [Hariharan R](#), [Hernandez O](#), [Hillier LW](#), [Holloway AK](#), [Janes D](#), [Janzen FJ](#), [Kandoth C](#), [Kong L](#), [de Koning AJ](#), [Li Y](#), [Litterman R](#), [McGaugh SE](#), [Mork L](#), [O'Laughlin M](#), [Paiz RT](#), [Pollock DD](#), [Ponting CP](#), [Radhakrishnan S](#), [Ranev BJ](#), [Richman JM](#), [St John J](#), [Schwartz T](#), [Sethuraman A](#), [Spinks PQ](#), [Storey KB](#), [Thane N](#), [Vinar T](#), [Zimmerman LM](#), [Warren WC](#), [Mardis ER](#), [Wilson RK](#).

Author information

Abstract

BACKGROUND: We describe the genome of the western painted turtle, *Chrysemys picta bellii*, one of the most widespread, abundant, and well-studied turtles. We place the genome into a comparative evolutionary context, and focus on genomic features associated with tooth loss, immune function, longevity, sex differentiation and determination, and the species' physiological capacities to withstand extreme anoxia and tissue freezing.

RESULTS: Our phylogenetic analyses confirm that turtles are the sister group to living archosaurs, and demonstrate an extraordinarily slow rate of evolution of the painted turtle to withstand complete anoxia and partial freezing appears to be associated with the ability to identify candidate genes for future functional analyses. Tooth loss shares a common pattern of evolution with birds, although the rate of accumulation of tooth loss genes generally reflect phylogeny rather than convergence. The genome shows signatures of strong natural selection, indicating that the painted turtle has evolved unique adaptations to survive in its environment.



Painted turtles:

- Adults endure extreme anoxia
- Hatchlings are freeze tolerant



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D. Hittel
Y. Maistrovski
S. Kornfeld
S. Alvarado
M. Chen
J.M. Storey

Funded by NSERC Canada



www.carleton.ca/~kbstorey



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Chemistry Bldg

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Click on photo



Vertebrate Freeze
Tolerance



Invertebrate Cold
Hardiness



Estivation



Hibernation



Anoxia tolerance

★ In the News!

2011 Fry Award, Canadian Society
of Zoologists [link](#), [link](#), [Lecture](#)
2010 Flavell Medalist, Royal Society
of Canada [link](#)

TEMPERATURE ADAPTATION IN
A CHANGING CLIMATE
eds: K.B. Storey & K. Tanino, 2012
& 600th publication from the Storey lab



[LINK](#)
Découverte [video](#) (français), March 2007
CBC [news](#), Jan. 2007
Discovery Channel [video](#), Jan. 2007
Discovery Channel [video](#), May 2006

[MEDIA](#): books, magazines, newspapers



Canada Research Chair Tier I
[PHOTO](#): Ken's [profile](#)

THE LAB

[Storey lab research Interests](#)

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[PEOPLE](#) in the Storey lab, Past and Present

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[Publications 1986 - 1995](#)

[Publications 1974 - 1985](#)

[MEDIA](#): Video & Radio interviews, Magazine & Newspaper articles, Textbook and Fiction features

BOOKS

[Temperature Adaptation in a Changing Climate](#):
CABI Publishers, 2012

[Cell and Molecular Responses to Stress](#)
Elsevier Science, 3 volumes, 2000-2002

[Environmental Stress and Gene Regulation](#)
BIOS Scientific Publishers, 1999

[Functional Metabolism: Regulation and
Adaptation](#)
John Wiley & Sons, 2004

[Molecular Mechanisms of Metabolic Arrest](#)
BIOS Scientific Publishers, 2001